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(54) Title: USE OF HEPARANASE TO IDENTIFY AND ISOLATE ANTI-HEPARANASE COMPOUND

(57) Abstract

Purified heparanase having activity of greater than 20 units/μg protein, preferably greater than 50 units heparanase activity per μg protein, is described. The use of heparanase for screening for anti-heparanase compounds is also described. In addition, the use of the high potency heparanase to accelerate wound healing or its use as an immobilized heparanase filter connected to extracorporeal devices to degrade heparin and neutralize its anticoagulant properties during surgery is disclosed.

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USE OF HEPARANASE TO IDENTIFY AND ISOLATE ANTI-HEPARANASE COMPOUND

FIELD OF INVENTION

The present invention discloses the use of mammalian heparanase, preferably recombinant heparanase, for screening for anti-heparanase compounds. More particularly, the present invention provides a method of selecting IHA (Inhibitors of Heparanase Activity). In addition, the present invention provides a purified heparanase, particularly suitable for use to identify and isolate anti-heparanase compounds as well as for other known uses of heparanases, such as its use to accelerate wound healing or its use as an immobilized heparanase filter connected to extracorporeal devices to degrade heparin and neutralize its anticoagulant properties during surgery.

BACKGROUND OF THE INVENTION

Elevated heparanase activity has been documented in mobile, invasive cells. Examples include; invasive melanoma, lymphoma, mastocytoma, mammary adenocarcinoma, leukemia, and rheumatoid fibroblasts. This activity has also been documented in non-pathologic situations involving the migration of lymphocytes, neutrophils, macrophages, eosinphils and platelets. An inhibitor of heparanase would therefore broadly influence the invasive potential of these diverse cells.

Inhibition of heparan sulfate degradation would also inhibit the release of bound growth factors and other biologic response modifiers that would, if released, fuel the growth of adjacent tissues and provide a supportive environment for cell growth (Rapraeger, et al., *Science* **252:** 1705-1708, 1991). Inhibitors of heparanase activity would be of value in the treatment of arthritis, vascular restenosis, tumor growth and progression, and fibro-proliferative disorders.

Until now, the obstacles to designing a screening assay to find inhibitors of mammalian heparanase have been the unavailability of a mammalian heparanase that is purified to apparent homogeneity and the lack of information about the amino acid sequence or the 3-dimensional structure of the enzyme. Without the amino acid sequence, it has not been possible to produce recombinant mammalian heparanase to be used in large volume screening efforts. Knowledge of the tertiary and quaternary structures would facilitate rational design of IHA. This report overcomes obstacles relating to the sequence of the heparanase, and also provides a model for higher-order structure.

Heparanase refers to a mammalian enzyme which can degrade heparin proteoglycans (HPG) and/or heparan sulfate proteoglycans (HSPG).

Heparanase activity in mammalian cells is well known. It is found in various melanoma cells (Nakajima, *et al., Cancer Letters* **31:** 277-283, 1986), mammary adenocarcinoma cells (Parish, *et al., Int. J. Cancer*, **40:** 511-518, 1987), leukemic cells (Yahalom, *et al., Leukemia*

Research **12:** 711-717, 1988), mast cells (Ogren and Lindahl, *J. Biol. Chem.* **250:** 2690-2697, 1975), macrophages (Savion, *et al., J. Cell. Physiol.*, **130:** 85-92, 1987), mononuclear cells (Sewell, *et al., Biochem. J.* **264:** 777-783, 1989), neutrophils (Matzner, *et al., J. Leukocyte Biology* **51:** 519-524, 1992), T-cells (Vettel, *et al., Eur. J. Immunol.* **21:** 2247-2251, 1991), platelets (Haimovitz-Friedman, *et al., Blood* **78:** 789-796, 1991), endothelial cells (Godder, *et al., J. Cell Physiol.* **148:** 274-280, 1991), and placenta (Klein and von Figura, *BBRC* **73:** 569, 1976).

WO 91/02977, incorporated herein by reference, describes a substantially, but partially, purified heparanase produced by cation exchange resin chromatography and the affinity absorbent purification of heparanase-containing cell extract. WO 91/02977 also describes a method promoting wound healing utilizing compositions comprising a "purified" form of heparanase.

Others have proposed the use of purified bacterial heparanase, immobilized onto filters and connected to extracorporeal devices, to degrade heparin and neutralize its anticoagulant properties post surgery (Freed, *et al., Ann. Biomed. Eng.* **21:** 67-76, 1993).

U.S. Patent 4,882,318 describes heparanase-inhibiting compositions for preventing tumor metastasis.

Haimovitz-Friedman *et al.* (*Blood* **78:** 789-796, 1991) describe an assay for heparanase activity that involves the culturing of endothelial cells in radiolabeled $^{35}SO_4$ to produce radiolabeled heparan sulfate proteoglycans, the removal of the cells which leaves the deposited extracellular matrix that contains the ^{35}S-HSPG, the addition of potential sources of heparanase activity, and the detection of possible activity by passing the supernatant from the radiolabeled extracellular matrix over a gel filtration column and monitoring for changes of the size of the radiolabeled material that would indicate that HSPG degradation had taken place. This assay does not have the capability for large-scale screening of inhibitors.

Nakajima *et al.* (*Anal. Biochem.* **196:** 162-171, 1986) describe a solid-phase substrate for the assay of melanoma heparanase activity. Heparan sulfate from bovine lung is chemically radiolabeled by reacting it with [^{14}C]-acetic anhydride. Free amino groups of the [^{14}C]-heparan sulfate were acetylated and the reducing termini were aminated. The [^{14}C]-heparan sulfate was chemically coupled to an agarose support via the introduced amine groups on the reducing termini. This substrate is limited in that it is an extensively chemically modified form of naturally occurring heparan sulfate.

Khan and Newman (*Anal. Biochem.* **196:** 373-376, 1991) describe an indirect assay for heparanase activity. In this assay, heparin is quantitated by its ability to interfere with the color development between a protein and the dye Coomassie brilliant blue. Heparanase activity is

detected by the loss of this interference. This assay is limited in use for screening because it is so indirect that other non-heparin compounds could also interfere with the protein-dye reaction.

The CXC chemokine family (also called the intercrine α family) is one branch of the supergene "intercrine" cytokine family (Oppenheim, *Ann. Rev. Biochem.* **9**: 617-648, 1991). It's members include platelet factor 4, platelet basic protein and derivatives, γIP-10, gro(α,β,γ), NAP-1/interleukin-8, mig, and ENA-78 (for review, see Miller and Krangel, *Critical Reviews in Immunology* **12**: 17-46, 1992). The other branch, the CC chemokines or intercrine-β family, includes MIP1α, MIP1β,JE/MCP-1, RANTES, and MCAF. All members of both branches of this chemokine family characteristically are basic heparin-binding polypeptides, display molecular weights between 8 and 11 kD, share 20 - 50% homology, and function broadly in pathologic situations characterized by inflammation and tissue remodeling.

The proteolytically processed forms of platelet basic protein include CTAP-III, β-thromboglobulin, and NAP-2. β-thromboglobulin (Moore, *et al.*, *Biochim. Biophys. Acta.* **379**: 360-369, 1975) and CTAP-III (Castor, *et al.*, *Arthritis Rheum.* **20**: 859-868, 1977), were originally isolated from activated supernatants or lysates from outdated platelets. Using the techniques of subcellular fractionation and radioimmunoassay, β-thromboglobulin was identified as an α-granule protein that could be released upon activation (Kaplan, *et al.*, *Blood* **53**: 604-618, 1979). Platelet basic protein itself was later isolated from fresh platelets, megakaryocytes, and HEL cells, an immortal human erythroleukemia cell line (Holt, *et al.*, *Biochemistry* **25**: 1988-1996, 1986; Holt, *et al.*, *Exp. Hematol.* **16**: 302-306, 1988). Walz and Baggiolini isolated the processed form of NAP-2 from platelet-containing cultures of stimulated mononuclear cells (Walz, *et al.*, *J. Exp. Med.* **170**: 1745-1750, 1989).

Material labeled as β-thromboglobulin is commercially available from Calbiochem, San Diego, CA (Cat. # 605165), Celsus Laboratories, Cincinnati, OH (Cat. # 41705), and Haematologic Technologies, Essex Jct., VT (Cat. # HBTG-02100. The inventors have determined, by using the "Purification Assay," that the commercial preparation have heparanase activity at a level of 0.075 units/μg. This activity is below the level of 1 unit/μg needed for the screening of anti-heparanase compounds in accordance with the assay of the subject invention.

U.S. Patent 4,897,348 describes recombinant materials and methods for producing human connective tissue-activating peptide-III (CTAP-III) and analogs thereof.

Transglutaminases catalyze the posttranslational modification of proteins by transamidation of available glutamine residues. This action results primarily in the formation of epsilon-(gamma-glutamyl)lysine cross-links (Greenberg, *et al.*, *FASEB J.* **5**: 3071-3076, 1991). This posttranslational modification has been reported to dramatically alter the action of some small proteins. For example, a transglutaminase produces a glutamine-lysine cross-link in the 13

kD phospholipase A_2 and increases its specific enzymatic activity (Cordella-Miele, *et al., J. Biol. Chem.* **265:** 17180-17188, 1990). A transglutaminase cross-links another small molecule, interleukin-2, and converts its activity to one that is cytotoxic to mature oligodendrocytes (Eitan and Schwartz, *Science* **261:** 106-108,1993). The glutamine-lysine cross-link in a protein would result in the loss of overall positive charge for that protein. The transglutaminases are optimally active and generally used under reducing conditions such as dithiothreitol. The concept that glutamine-lysine cross-linking alters the activity of these small proteins may be applicable to other small molecules as well.

SUMMARY OF THE INVENTION

The present invention discloses a method of screening for compounds having anti-heparanase activity (AHA compounds), i.e. inhibitors of heparanase activity (IHA), comprising the steps of: contacting a potential AHA compound with radiolabeled heparin/heparan sulfate and heparanase for a time and under such conditions sufficient to allow for inhibition of heparanase activity; detecting inhibition of heparanase activity; and selecting compounds that inhibit heparanase activity. The present invention also discloses the amino acid sequence identity of the heparanase that has been purified to homogeneity by chromatography under reducing conditions. Identification of the amino acid sequence of the protein which contains heparanase activity is crucial for the production of recombinant mammalian heparanase.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides a purified heparanase, and a method for producing it. The heparanase so produced has an activity of greater than 20 units/ µg protein, preferably greater than 50 units heparanase activity per µg protein (1 unit = 1% cpm < 30 K/hr using the "Purification Assay" (Example 2, Part D).

In addition, the present invention provides recombinant heparanase and a means for producing it. The term "purified heparanase" as used in the specification and claims includes the recombinant heparanase as described in the subject application. The recombinant heparanase of the subject invention can be used for the same purposes and in the same manner as the purified heparanase.

The purified heparanase of the present invention has an isoelectric point of less than 5.5 (preferably about 4.8 - 5.1) and preferably is activated by treatment with transglutaminase using reducing conditions.

The recombinant heparanase of the present invention has an isoelectric point of less than 5.5 (preferably about 4.8 - 5.1), and is isolated under reducing conditions and is activated by treatment with transglutaminase.

Suitable transglutaminases that may be used for this purpose include Activated Factor XIIIa, guinea pig liver transglutaminase, epidermal transglutaminase, keratinocyte

transglutaminase, and tissue transglutaminase.

The heparanase of the present invention has the amino acid sequence (SEQ. ID. NO: 1) of:

Asn Leu Ala Lys Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Tyr Ala
5 10 15

Glu Leu Arg Cys Met Cys Ile Lys Thr Thr Ser Gly Ile His Pro Lys
20 25 30

Asn Ile Gln Ser Leu Glu Val Ile Gly Lys Gly Thr His Cys Asn Gln
35 40 45

Val Glu Val Ile Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp
50 55 60

Pro Asp Ala Pro Arg Ile Lys Lys Ile Val Gln Lys Lys Leu Ala Gly
65 70 75 80

Asp Glu Ser Ala Asp
85

encoded by the cDNA sequence (SEQ ID NO: 2) of:

1 AACTTGGCGA AAGGCAAAGA GGAAAGTCTA GACAGTGACT TGTATGCTGA

51 ACTCCGCTGC ATGTGTATAA AGACAACCTC TGGAATTCAT CCCAAAAACA

101 TCCAAAGTTT GGAAGTGATC GGGAAAGGAA CCCATTGCAA CCAAGTCGAA

151 GTGATAGCCA CACTGAAGGA TGGGAGGAAA ATCTGCCTGG ACCCAGATGC

201 TCCCAGAATC AAGAAAATTG TACAGAAAAA ATTGGCAGGT GATGAATCTG

251 CTGAT

which corresponds to the cDNA sequence and derived amino acid sequence of CTAP-III. See Wenger et al., *Blood*, 73: 1498-1503, 1989.

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 3) of:

Ser Ser Thr Lys Gly Gln Thr Lys Arg Asn Leu Ala Lys Gly Lys Glu
5 10 15

Glu Ser Leu Asp Ser Asp Leu Tyr Ala Glu Leu Arg Cys Met Cys Ile
20 25 30

Lys Thr Thr Ser Gly Ile His Pro Lys Asn Ile Gln Ser Leu Glu Val
35 40 45

Ile Gly Lys Gly Thr His Cys Asn Gln Val Glu Val Ile Ala Thr Leu
50 55 60

Lys Asp Gly Arg Lys Ile Cys Leu Asp Pro Asp Ala Pro Arg Ile Lys
65 70 75 80

Lys Ile Val Gln Lys Lys Leu Ala Gly Asp Glu Ser Ala Asp
85 90

encoded by the cDNA sequence (SEQ ID NO: 4) of:

1 TCCTCCACCA AAGGACAAAC TAAGAGAAAC TTGGCGAAAG GCAAAGAGGA
51 AAGTCTAGAC AGTGACTTGT ATGCTGAACT CCGCTGCATG TGTATAAAGA
101 CAACCTCTGG AATTCATCCC AAAAACATCC AAAGTTTGGA AGTGATCGGG
151 AAAGGAACCC ATTGCAACCA AGTCGAAGTG ATAGCCACAC TGAAGGATGG
201 GAGGAAAATC TGCCTGGACC CAGATGCTCC CAGAATCAAG AAAATTGTAC
251 AGAAAAAATT GGCAGGTGAT GAATCTGCTG AT

which corresponds to the cDNA sequence and derived amino acid sequence of platelet basic protein. See Wenger et al., *Blood*, 73: 1498-1503, 1989 as well as Walz and Baggiolini, *BBRC* 159: 969-981, 1989; Castor, *et al.*, *BBRC* 163: 1071-1078, 1989.

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 5) of:

Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Tyr Ala Glu Leu Arg Cys
1 5 10 15

Met Cys Ile Lys Thr Thr Ser Gly Ile His Pro Lys Asn Ile Gln Ser
20 25 30

Leu Glu Val Ile Gly Lys Gly Thr His Cys Asn Gln Val Glu Val Ile
35 40 45

Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp Pro Asp Ala Pro
50 55 60

Arg Ile Lys Lys Ile Val Gln Lys Lys Leu Ala Gly Asp Glu Ser Ala
65 70 75 80

Asp

encoded by the cDNA sequence (SEQ ID NO: 6) of:

1 GGCAAAGAGG AAAGTCTAGA CAGTGACTTG TATGCTGAAC TCCGCTGCAT
51 GTGTATAAAG ACAACCTCTG GAATTCATCC CAAAAACATC CAAAGTTTGG
101 AAGTGATCGG GAAAGGAACC CATTGCAACC AAGTCGAAGT GATAGCCACA
151 CTGAAGGATG GGAGGAAAAT CTGCCTGGAC CCAGATGCTC CCAGAATCAA
201 GAAAATTGTA CAGAAAAAAT TGGCAGGTGA TGAATCTGCT GAT

which corresponds to the cDNA sequence and derived amino acid sequence β-thromboglobulin (βTG). See Wenger et al., *Blood*, 73: 1498-1503, 1989.

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 7) of:

Glu Leu Arg Cys Met Cys Ile Lys Thr Thr Ser Gly Ile His Pro Lys
1 5 10 15

Asn Ile Gln Ser Leu Glu Val Ile Gly Lys Gly Thr His Cys Asn Gln
20 25 30

Val Glu Val Ile Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp
35 40 45

Pro Asp Ala Pro Arg Ile Lys Lys Ile Val Gln Lys Lys Leu Ala Gly
50 55 60

Asp Glu Ser Ala Asp
65

encoded by the cDNA sequence (SEQ ID NO: 8) of:

1 GAACTCCGCT GCATGTGTAT AAAGACAACC TCTGGAATTC ATCCCAAAAA
51 CATCCAAAGT TTGGAAGTGA TCGGGAAAGG AACCCATTGC AACCAAGTCG
101 AAGTGATAGC CACACTGAAG GATGGGAGGA AAATCTGCCT GGACCCAGAT
151 GCTCCCAGAA TCAAGAAAAT TGTACAGAAA AAATTGGCAG GTGATGAATC
201 TGCTGAT

which corresponds to the cDNA sequence and derived amino acid sequence of neutrophil activating peptide-2.

The foregoing amino acid sequences correspond to the products of a single gene called platelet basic protein (Walz and Baggiolini, *BBRC* 159: 969-981, 1989; Castor, *et al.*, *BBRC* 163: 1071-1078, 1989). The complete gene sequence of platelet basic protein is well known. See, for example, Wenger et al., *Blood*, 73: 1498-1503, 1989 and *Proc. Natl. Acad. Sci. USA*, 90, 3660-3664, 1993.

The present invention also provides heparanase having the amino acid sequences of other members of the CXC chemokine family [including Platelet factor 4 (SEQ. ID NO. 12), γIP-10 (SEQ. ID NO. 14), *gro*/MGSA (SEQ. ID NO. 16), *gro*-β/MIP-2α (SEQ. ID NO. 18), *gro*-γ/MIP-2β (SEQ. ID NO. 20), Interleukin-8/NAP-1 (SEQ. ID NO. 22) and ENA-78 (SEQ. ID NO. 24)] as well as members of the CC chemokine family [including MIP-1α (SEQ. ID NO. 26), MIP-1β (SEQ. ID NO. 28), I-309 (SEQ. ID NO. 23), MCP-1 (SEQ. ID NO. 32), MCP-3 (SEQ. ID NO. 34), RANTES (SEQ. ID NO. 36), fic (SEQ. ID NO. 38) and MCP-2 (SEQ. ID NO. 40)]; purified to apparent homogeneity, prepared in the presence of reducing conditions, and activated by treatment with transglutaminase. Suitable transglutaminases that may be used for this purpose include Activated Factor XIIIa, guinea pig liver transglutaminase, epidermal transglutaminase, keratinocyte transglutaminase, and tissue transglutaminase.

In another aspect, the present invention provides a heparanase having the amino acid

sequence (SEQ ID NO: 12) of:

Met Ser Ser Ala Ala Gly Phe Cys Ala Ser Arg Pro Gly Leu Leu Phe Leu Gly Leu Leu Leu Leu Pro Leu Val Val Ala Phe Ala Ser Ala Glu Ala Glu Glu Asp Gly Asp Leu Gln Cys Leu Cys Val Lys Thr Thr Ser Gln Val Arg Pro Arg His Ile Thr Ser Leu Glu Val Ile Lys Ala Gly Pro His Cys Pro Thr Ala Gln Leu Ile Ala Thr Leu Lys Asn Gly Arg Lys Ile Cys Leu Asp Leu Gln Ala Pro Leu Tyr Lys Lys Ile Ile Lys Lys Leu Leu Glu Ser

encoded by the cDNA sequence (SEQ ID NO: 13) of:

1 CCGCAGCATG AGCTCCGCAG CCGGGTTCTG CGCCTCACGC CCCGGGCTGC
51 TGTTCCTGGG GTTGCTGCTC CTGCCACTTG TGGTCGCCTT CGCCAGCGCT
101 GAAGCTGAAG AAGATGGGGA CCTGCAGTGC CTGTGTGTGA AGACCACCTC
151 CCAGGTCCGT CCCAGGCACA TCACCAGCCT GGAGGTGATC AAGGCCGGAC
201 CCCACTGCCC CACTGCCCAA CTGATAGCCA CGCTGAAGAA TGGAAGGAAA
251 ATTTGCTTGG ACCTGCAAGC CCCGCTGTAC AAGAAAATAA TTAAGAAACT
301 TTTGGAGAGT TAGCTACTAG CTGCCTACGT GTGTGCATTT GCTATATAGC
351 ATACTTCTTT TTTCCAGTTT CAATCTAACT GTGAAAGAAA CTTCTGATAT
401 TTGTGTTATC CTTATGATTT TAAATAAACA AAATAAATC

which corresponds to the cDNA sequence and derived amino acid sequence of platelet factor 4. See Poncz et al., *Blood* 69, 219-223 (1987).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 14) of:

Met Asn Gln Thr Ala Ile Leu Ile Cys Cys Leu Ile Phe Leu Thr Leu Ser Gly Ile Gln Gly Val Pro Leu Ser Arg Thr Val Arg Cys Thr Cys Ile Ser Ile Ser Asn Gln Pro Val Asn Pro Val Asn Pro Arg Ser Leu Glu Lys Leu Glu Ile Ile Pro Ala Ser Gln Phe Cys Pro Arg Val Glu Ile Ile Ala Thr Met Lys Lys Lys Gly Glu Lys Arg Cys Leu Asn Pro Glu Ser Lys Ala Ile Lys Asn Leu Leu Lys Ala Val Ser Lys Glu Met Ser Lys Arg Ser Pro

encoded by the cDNA sequence (SEQ ID NO: 15) of:

1 GAGACATTCC TCAATTGCTT AGACATATTC TGAGCCTACA GCAGAGGAAC
51 CTCCAGTCTC AGCACCATGA ATCAAACTGC GATTCTGATT TGCTGCCTTA
101 TCTTTCTGAC TCTAAGTGGC ATTCAAGGAG TACCTCTCTC TAGAACCGTA
151 CGCTGTACCT GCATCAGCAT TAGTAATCAA CCTGTTAATC CAAGGTCTTT
201 AGAAAAACTT GAAATTATTC CTGCAAGCCA ATTTTGTCCA CGTGTTGAGA
251 TCATTGCTAC AATGAAAAAG AAGGGTGAGA AGAGATGTCT GAATCCAGAA
301 TCGAAGGCCA TCAAGAATTT ACTGAAAGCA GTTAGCAAGG AAATGTCTAA
351 AAGATCTCCT TAAAACCAGA GGGGAGCAAA ATCGATGCAG TGCTTCCAAG
401 GATGGACCAC ACAGAGGCTG CCTCTCCCAT CACTTCCCTA CATGGAGTAT

451 ATGTCAAGCC ATAATTGTTC TTAGTTTGCA GTTACACTAA AAGGTGACCA
501 ATGATGGTCA CCAAATCAGC TGCTACTACT CCTGTAGGAA GGTTAATGTT
551 CATCATCCTA AGCTATTCAG TAATAACTCT ACCCTGGCAC TATAATGTAA
601 GCTCTACTGA GGTGCTATGT TCTTAGTGGA TGTTCTGACC CTGCTTCAAA

which corresponds to the cDNA sequence and derived amino acid sequence γIP-10. See Luster et al., Nature 315, 672-676 (1985).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 16) of:

Met Ala Arg Ala Ala Leu Ser Ala Ala Pro Ser Asn Pro Arg Leu Leu Arg Val Ala Leu
Leu Leu Leu Leu Leu Val Ala Ala Gly Arg Arg Ala Ala Gly Ala Ser Val Ala Thr Glu
Leu Arg Cys Gln Cys Leu Gln Thr Leu Gln Gly Ile His Pro Lys Asn Ile Gln Ser Val
Asn Val Lys Ser Pro Gly Pro His Cys Ala Gln Thr Glu Val Ile Ala Thr Leu Lys Asn
Gly Arg Lys Ala Cys Leu Asn Pro Ala Ser Pro Ile Val Lys Lys Ile Ile Glu Lys Met
Leu Asn Ser Asp Lys Ser Asn

encoded by the cDNA sequence (SEQ ID NO: 17) of:

1 CTCGCCAGCT CTTCCGCTCC TCTCACAGCC GCCAGACCCG CCTGCTGAGC
51 CCCATGGCCC GCGCTGCTCT CTCCGCCGCC CCCAGCAATC CCCGGCTCCT
101 GCGAGTGGCA CTGCTGCTCC TGCTCCTGGT AGCCGCTGGC CGGCGCGCAG
151 CAGGAGCGTC CGTGGCCACT GAACTGCGCT GCCAGTGCTT GCAGACCCTG
201 CAGGGAATTC ACCCCAAGAA CATCCAAAGT GTGAACGTGA AGTCCCCCGG
251 ACCCCACTGC GCCCAAACCG AAGTCATAGC CACACTCAAG AATGGGCGGA
301 AAGCTTGCCT CAATCCTGCA TCCCCCATAG TTAAGAAAAT CATCGAAAAG
351 ATGCTGAACA GTGACAAATC CAACTGACCA GAAGGGAGGA GGAAGCTCAC
401 TGGTGGCTGT TCCTGAAGGA GGCCCTGCCC TTATAGGAAC AGAAGAGGAA
451 AGAGAGACAC AGCTGCAGAG GCCACCTGGA TTGTGCCTAA TGTGTTTGAG
501 CATCGCTTAG GAGAAGTCTT CTATTTATTT ATTTATTCAT TAGTTTTGAA
551 GATTCTATGT TAATATTTTA GGTGTAAAAT AATTAAGGGT ATGATTAACT
601 CTACCTGCAC ACTGTCCTAT TATATTCATT CTTTTTGAAA TGTCAACCCC
651 AAGTTAGTTC AATCTGGATT CATATTTAAT TTGAAGGTAG AATGTTTTCA
701 AATGTTCTCC AGTCATTATG TTAATATTTC TGAGGAGCCT GCAACATGCC
751 AGCCACTGTG ATAGAGGCTG GCGGATCCAA GCAAATGGCC AATGAGATCA
801 TTGTGAAGGC AGGGGAATGT ATGTGCACAT CTGTTTTGTA ACTGTTTAGA
851 TGAATGTCAG TTGTTATTTA TTGAAATGAT TTCACAGTGT GTGGTCAACA
901 TTTCTCATGT TGAAACTTTA AGAACTAAAA TGTTCTAAAT ATCCCTTGGA
951 CATTTTATGT CTTTCTTGTA AGGCATACTG CCTTGTTTAA TGGTAGTTTT
1001 ACAGTGTTTC TGGCTTAGAA CAAAGGGGCT TAATTATTGA TGTTTTCGGA

which corresponds to the cDNA sequence and derived amino acid sequence of *gro*/MGSA (melanoma growth stimulatory activity). See Anisowicz et al., Proc. Natl. Acad. Sci. U.S.A. 84, 7188-7192 (1987).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 18) of:

Met Ala Arg Ala Thr Leu Ser Ala Ala Pro Ser Asn Pro Arg Leu Leu Arg Val Ala Leu Leu Leu Leu Leu Leu Val Ala Ala Ser Arg Arg Ala Ala Gly Ala Pro Lys Ala Thr Glu Lys Arg Cys Gln Cys Lys Gln Thr Leu Gln Gly Ile His Leu Lys Asn Ile Gln Ser Val Lys Val Lys Ser Pro Gly Pro His Cys Ala Gln Thr Glu Val Ile Ala Thr Leu Lys Asn Gly Gln Lys Ala Cys Leu Asn Pro Ala Ser Pro Met Val Lys Lys Ile Ile Glu Lys Met Leu Lys

encoded by the cDNA sequence (SEQ ID NO: 19) of:

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   1 CTCTCCTCCT CGCACAGCCG CTCGAACCGC CTGCTGAGCC CCATGGCCCG
  51 CGCCACGCTC TCCGCCGCCC CCAGCAATCC CCGGCTCCTG CGGGTGGCGC
 101 TGCTGCTCCT GCTCCTGGTG GCCGCCAGCC GGCGCGCAGC AGGAGCGCCC
 151 CTGGCCACTG AACTGCGCTG CCAGTGCTTG CAGACCCTGC AGGGAATTCA
 201 CCTCAAGAAC ATCCAAAGTG TGAAGGTGAA GTCCCCCGGA CCCCACTGCG
 251 CCCAAACCGA AGTCATAGCC ACACTCAAGA ATGGGCAGAA AGCTTGTCTC
 301 AACCCCGCAT CGCCCATGGT TAAGAAAATC ATCGAAAAGA TGCTGAAAAA
 351 TGGCAAATCC AACTGACCAG AAGGAAGGAG GAAGCTTATT GGTGGCTGTT
 401 CCTGAAGGAG GCCCTGCCCT TACAGGAACA GAAGAGGAAA GAGAGACACA
 451 GCTGCAGAGG CCACCTGGAT TGCGCCTAAT GTGTTTGAGC ATCACTTAGG
 501 AGAAGTCTTC TATTTATTTA TTTATTTATT TATTTGTTTG TTTTAGAAGA
 551 TTCTATGTTA ATATTTTATG TGTAAAATAA GGTTATGATT GAATCTACTT
 601 GCACACTCTC CCATTATATT TATTGTTTAT TTTAGGTCAA ACCCAAGTTA
 651 GTTCAATCCT GATTCATATT TAATTTGAAG ATAGAAGGTT TGCAGATATT
 701 CTCTAGTCAT TTGTTAATAT TTCTTCGTGA TGACATATCA CATGTCAGCC
 751 ACTGTGATAG AGGCTGAGGA ATCCAAGAAA ATGGCCAGTG AGATCAATGT
 801 GACGGCAGGG AAATGTATGT GTGTCTATTT TGTAACTGTA AAGATGAATG
 851 TCAGTTGTTA TTTATTGAAA TGATTTCACA GTGTGTGGTC AACATTTCTC
 901 ATGTTGAAGC TTTAAGAACT AAAATGTTCT AAATATCCCT TGGACATTTT
 951 ATGTCTTTCT TGTAAGGCAT ACTGCCTTGT TTAATGTTAA TTATGCAGTG
1001 TTTCCCTCTG TGTTAGAGCA GAGAGGTTTC GATATTTATT GATGTTTTCA
1051 CAAAGAACAG GAAAATAAAA TATTTAAAAA T
```

which corresponds to the cDNA sequence and derived amino acid sequence *gro*-β/MIP-2α (macrophage inflammatory protein 2-α). See Tekamp-Olson et al., J. Exp. Med. 172, 911-919

(1990).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 20) of:

Met Ala His Ala Thr Leu Ser Ala Ala Pro Ser Asn Pro Arg Leu Leu Arg Val Ala Leu Leu Leu Leu Leu Leu Val Ala Ala Ser Arg Arg Ala Ala Gly Ala Ser Val Val Thr Glu Leu Arg Cys Gln Cys Leu Gln Thr Leu Gln Gly Ile His Leu Lys Asn Ile Gln Ser Val Asn Val Arg Ser Pro Gly Pro His Cys Ala Gln Thr Glu Val Ile Ala Thr Leu Lys Asn Gly Lys Lys Ala Cys Leu Asn Pro Ala Ser Pro Met Val Gln Lys Ile Ile Glu Lys Ile Leu Asn Lys Gly Ser Thr Asn

encoded by the cDNA sequence (SEQ ID NO: 21) of:

1 CTCGCACAGC TTCCCGACGC GTCTGCTGAG CCCCATGGCC CACGCCACGC
51 TCTCCGCCGC CCCCAGCAAT CCCCGGCTCC TGCGGGTGGC GCTGCTGCTC
101 CTGCTCCTGG TGGCCGCCAG CCGGCGCGCA GCAGGAGCGT CCGTGGTCAC
151 TGAACTGCGC TGCCAGTGCT TGCAGACACT GCAGGGAATT CACCTCAAGA
201 ACATCCAAAG TGTGAATGTA AGGTCCCCCG GACCCCACTG CGCCCAAACC
251 GAAGTCATAG CCACACTCAA GAATGGGAAG AAAGCTTGTC TCAACCCCGC
301 ATCCCCCATG GTTCAGAAAA TCATCGAAAA GATACTGAAC AAGGGGAGCA
351 CCAACTGACA GGAGAGAAGT AAGAAGCTTA TCAGCGTATC ATTGACACTT
401 CCTGCAGGGT GGTCCCTGCC CTTACCAGAG CTGAAAATGA AAAAGAGAAC
451 AGCAGCTTTC TAGGGACAGC TGGAAAGGAC TTAATGTGTT TGACTATTTC
501 TTACGAGGGT TCTACTTATT TATGTATTTA TTTTTGAAAG CTTGTATTTT
551 AATATTTTAC ATGCTGTTAT TTAAAGATGT GAGTGTGTTT CATCAAACAT
601 AGCTCAGTCC TGATTATTTA ATTGGAATAT GATGGGTTTT AAATGTGTCA
651 TTAAACTAAT ATTTAGTGGG AGACCATAAT GTGTCAGCCA CCTTGATAAA
701 TGACAGGGTG GGGAACTGGA GGGTGGGGGG ATTGAAATGC AAGCAATTAG
751 TGGATCACTG TTAGGGTAAG GGAATGTATG TACACATCTA TTTTTTATAC
801 TTTTTTTTTA AAAAAAGAAT GTCAGTTGTT ATTTATTCAA ATTATCTCAC
851 ATTATGTGTT CAACATTTTT ATGCTGAAGT TTCCCTTAGA CATTTTATGT
901 CTTGCTTGTA GGGCATAATG CCTTGTTTAA TGTCCATTCT GCAGCGTTTC
951 TCTTTCCCTT GGAAAAGAGA ATTTATCATT ACTGTTAC

which corresponds to the cDNA sequence and derived amino acid sequence *gro*-γ/MIP-2β (macrophage inflammatory protein 2-β). See Tekamp-Olson et al., J. Exp. Med. 172, 911-919 (1990).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 22) of:

Met Thr Ser Lys Leu Ala Val Ala Leu Leu Ala Ala Phe Leu Ile Ser Ala Ala Leu Cys

Glu Gly Ala Val Leu Pro Arg Ser Ala Lys Glu Leu Arg Cys Gln Cys Ile Lys Thr Tyr Ser Lys Pro Phe His Pro Lys Phe Ile Lys Glu Leu Arg Val Ile Glu Ser Gly Pro His Cys Ala Asn Thr Glu Ile Ile Val Lys Leu Ser Asp Gly Arg Glu Leu Cys Leu Asp Pro Lys Glu Asn Trp Val Gln Arg Val Val Glu Lys Phe Leu Lys Arg Ala Glu

encoded by the cDNA sequence (SEQ ID NO: 23) of:

```
  1 ATGACTTCCA AGCTGGCCGT GGCTCTCTTG GCAGCCTTCC TGATTTCTGC
 51 AGCTCTGTGT GAAGGTGCAG TTTTGCCAAG GAGTGCTAAA GAACTTAGAT
101 GTCAGTGCAT AAAGACATAC TCCAAACCTT TCCACCCCAA ATTTATCAAA
151 GAACTGAGAG TGATTGAGAG TGGACCACAC TGCGCCAACA CAGAAATTAT
201 TGTAAAGCTT TCTGATGGAA GAGAGCTCTG TCTGGACCCC AAGGAAAACT
251 GGGTGCAGAG GGTTGTGGAG AAGTTTTTGA AGAGGGCTGA G
```

which corresponds to the cDNA sequence and derived amino acid sequence Interleukin-8/NAP-1 (neutrophil activating protein-1). See Kunser et al., Kidney Int. 39, 1240-1248 (1991).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 24) of:

Ala Gly Pro Ala Ala Ala Val Leu Arg Glu Lys Arg Cys Val Cys Leu Gln Thr Thr Gln Gly Val His Pro Lys Met Ile Ser Asn Leu Gln Val Phe Ala Ile Gly Pro Gln Cys Ser Lys Val Glu Val Val Ala Ser Leu Lys Asn Gly Lys Glu Ile Cys Leu Asp Pro Glu Ala Pro Phe Leu Lys Lys Val Ile Gln Lys Ile Leu Asp Gly Gly Asn Lys Glu Asn

encoded by the cDNA sequence (SEQ ID NO: 25) of:

```
  1 GTGTTGCGGG AACTGCGGTG CGTGTGTTTA CAGACCACGC AGGGAGTTCA
 51 TCCCAAAATG ATCAGTAATC TGCAAGTGTT CGCCATAGGC CCACAGTGCT
101 CCAAGGTGGA AGTGGTAGCC TCCCTGAAGA ACGGGAAGGA AATTTGTCTT
151 GATCCAGAAG CCCCTTTTCT AAAGAAAGTC ATCCAGAAAA TCCTCGACGG
201 CGGCAACAAA GAAAAC
```

which corresponds to the cDNA sequence and derived amino acid sequence of a novel inflammatory peptide (ENA-78) with homology to interleukin 8. See Walz et al., J. Exp. Med. 174, 1355-1362 (1991).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 26) of:

Met Gln Val Ser Thr Ala Ala Leu Ala Val Leu Leu Cys Thr Met Ala Leu Cys Asn Gln Val Leu Ser Ala Pro Leu Ala Ala Asp Thr Pro Thr Ala Cys Cys Phe Ser Tyr Thr Ser Arg Gln Ile Pro Gln Asn Phe Ile Ala Asp Tyr Phe Glu Thr Ser Ser Gln Cys Ser Lys Pro Ser Val Ile Phe Leu Thr Lys Arg Gly Arg Gln Val Cys Ala Asp Pro Ser Glu Glu Trp Val Gln Lys Tyr Val Ser Asp Leu Glu Leu Ser Ala

encoded by the cDNA sequence (SEQ ID NO: 27) of:

```
   1 GAATTCAAGG CCTGTCCTGG TTTGGTCCCA ATTTACCTTT ATCATCCATA
  51 TTCACCCCCA CTGCTCTGCA GCTCCACTGA AGCACCCCCT CTTTCCTCTG
 101 AGCCACAATG TCACACCCAG GACTCTGCCT CAGCTGGGCC TCCACTGCCC
 151 ACCCATCTAT AGATGCCTAA ATCCCGGGCA GTTATCCAGA CACAACTAAA
 201 GTTCCATCCC TTCCATGAAG CCTTCCCCAA CCCTCTGGTG GAAGGTCACT
 251 TCTTCCTCAT GGGGTTCTGA GCTTTCATTT CTTTTTCTAC TAAGAGTTTT
 301 ACAATTACCT GTTCATACAC TCTACCTGCC CCCATGAGAC CAGGGGCATC
 351 TCAGAAACAA AGATCATTAA AACCAACTAA ATCTATTTCT CATTATAAAA
 401 TGAGATATGC TGATTGATTG CAAAATAATA AAATAACAAA GTATGGAAAA
 451 GAAAAAAAAA AGCATATAAT CTGGCTGAGA AGGTAGAGAC CCTTCCACAC
 501 CACTGAAATT ATGTGTTGAA AAGAATAAGG AAAAAACTGC TTCAGTTTGG
 551 CATTATTTAT GTAAGTATAG TATAGGATCC TTAAAATGGT TCAAAGAAAT
 601 GGGAAATCAA GACTTCATTT TGGCAAAGCC ATTGAACAGA AACTGTAGCA
 651 TATTTATCAG TAATTTCTTT CAGATTAAAC AACTGACAAC AACCCACTTT
 701 TCAACCAGTG ATGTTGGAAA TGTTTTAAAA CAAAATTAGT TCATAAATTT
 751 GTGGGTTGAC CAAGAAGGTA ATAAAGTCTC ACTAAATAAA ATGAGGAAAA
 801 TTCAGAAAAA GAAAAAAATA AGAAAATAAA TCACCCATGG ATCTAAGCAC
 851 TATTCATTCT TTAAGGCATG TATTTCCAAG CCTTTTAATT TTTTCATGCC
 901 TAGAGTTGGC ATGGCATATA TATATCTTTA TACAATTCTT CAAATTTTAT
 951 AGAATTTGTA TAATGTTTTA TCTTGCTTTT TTTTTAACCA CTGATGTTAT
1001 AAGCATATTT ATGCCACTTC ATTCACGTTA GAGACTTAAT AATAAAGGAT
1051 CTTGTGGATA ATTTATCATT CCCTGATAGA GAAAAATTTA GCTTTGCTTA
1101 TTTTAGAGTT ATAAATGATG CTGGGTCAGG TATCTTTATG TTTGAAGATG
1151 GCTCCATATT TGGGTTGTTT CCACAGAACT CTTTCCAGAA ATGCTTTTTC
1201 TAGGTTAATG GCTACACATA TTTCTAGGCA CCTGACATAC TGACACCCAC
1251 CTCTAAAGTA TTTTTATGAT CCACAACTAG CGTTTAACAC AGCGCCCCAG
1301 TCACTCCGAG ACTAATAAAT AGACAAATGA CTGAAACGTG ACCTCATGCT
1351 TTCTATTCCT CCAGCTTTCA TTGAGTTCCT TTCCTCTGGG AGGACTGGGG
1401 GTTGTCTAGC CCTCCACAGC ATCAGCCCAT TGACCCTATC CTTGTGGTTA
1451 TAGCAGCTGA GGAAGCAGAA TT  AGCTCT GTGGGAAGGA ATGGGGCTGG
1501 AGAGTTCATG CATAGACCAA TTCTTTTTTT TTTTTTTTTT TGAGATGGAG
1551 TTTCACTTTT GTTGCCCAGG CTGGAGTGCA ATGGCATGAT CTCAGCTCAC
1601 CACAGCCCCC ACCTCCTGGG TTCAAGCGAT TCTCCTGCCC TCAGCCTCCC
1651 GAGTAGCTGG GATTACAGGC ATGTGCCACC ACGCCTGACT ACTTTTGTAT
1701 TTTTAGTAGA GATGGAGTTT CTCTTTCTTG GTCAGGTTGG TCTCAAACTC
1751 CTGACCTCAG GTGATCTGCA GCCTCGGCCT CCAAAGTGTT GGGATTACAG
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1801 GTGTGAGCGA CCATGCCTGG CTGCATAGAC CAGTTCTTAT GAGAAGGGAT
1851 CAACTAAGAA TAGCCTTGGG TTGACACACA CCCCTCTTCA CACTCACAGG
1901 AGAAACCCCA TGAAGCTAGA ACCAGTCATG AGTTGAGAGC TGAGAGTTAG
1951 AGAGTAGCTC AGAGATGCTA TTCTTGGATA TCCTGAGCCC CTGTGGTCAC
2001 CAGGGACCCT GAGTTGTGCA ACACTCAGCA TGACAGCATC ACTACACTTA
2051 AAAATTTCCC TCCTCACCCC CAGATTCCAT TTCCCCATCC GCCAGGGCTG
2101 CCTATAAAGA GGAGAGATGG CTTCAGACAT CAGAAGGACG CAGGCAGCAA
2151 AGAGTAGTCA GTCCCTTCTT GGCTCTGCTG ACACTCGAGC CCACATTCCA
2201 TCACCTGCTC CCAATCATGC AGGTCTCCAC TGCTGCCCTT GCCGTCCTCC
2251 TCTGCACCAT GGCTCTCTGC AACCAGGTCC TCTCTGCACC ACGTGAGTCC
2301 ATGTTGTTGT TGTGGGTATC ACCACTCTCT GGCCATGGTT AGACCACATC
2351 AGTCTTTTTT TGTGGCGTGA GAGGCCCCGA AGAGAAAAGA AGGAAGTTCT
2401 TAAAGCGCTG CCAAACACCT TGGTCTTTTT CTTCACAACT TTTATTTTTA
2451 TCTCTAGAAG GGGTCTTAGC CCTCCTAGTC TCCAGGTATG AGAATCTAGG
2501 CAGGGGCAGG GGAGTTACAG TCCCTTGTAC AGATAGAAAA ACAGGGTTCA
2551 AAACGAATCA GTTTGCAAGA GGCAGAATCC AGGGCTGCTT ACTTCCCAGT
2601 GGGGTCTGTT CTTCACTCTC CAGCTCACCC TAGTCTCCCA GGAGCCCTGT
2651 CCCTTGGATG TCTTATGAGA GATGTCCAGG GCTTCTCTTG GGCTGGGGTA
2701 TGACTTCTTG AACCGACAAA ATTCCATGAA GAGAGCTAAG AGAACAGTCC
2751 ATTCAGGTAT CTGGATCACA TAGAGAAACA GAGAACCCAC TATGAAGAGT
2801 CAAGGGGAAA GAGGAATATA GACAGAAACA AAGAGACATT TCTCTGCAAA
2851 ACCCCCCAAA TGCCTTGCAG TCACTTGGTC TGAGCAAGCC TGCCCTCCTC
2901 AACCACTCAG GGATCAGAAG CTGCCTGGCC TTTTCTTCTG AGCTGTGACT
2951 TGGGCTTATT CTCTCCTTTC TCCGCAGTTG CTGCTGACAC GCCGACCGCC
3001 TGCTGCTTCA GCTACACCTC CCGACAGATT CCACAGAATT TCATAGCTGA
3051 CTACTTTGAG ACGAGCAGCC AGTGCTCCAA GCCCAGTGTC ATGTAAGTGC
3101 CAGTCTTCCT GCTCACCTCT AGGGAGGTAG GGAGTGTCAG GGTGGGGGCA
3151 GAAACAGGCC AGAAGGCCAT CCTGGAAAGG CCCAGCCTTC AGGAGCCTAT
3201 CGGGGATACA GGACGCAGGG CACTGAGGTG TGACCTGACT TGGGGCTGGA
3251 GTGAGGTGGG TGTTACAGAG TCAGGAAGGG CTGCCCCAGG CCAGAGGAAA
3301 GGGACAGGAA GAAGGAGGCA GCAGGACACT CTGAGGGCCC CCTTGCCTGG
3351 AGTCACTGAG AGAAGCTCTC TAGACGGAGA TAGGCAGGGG GCCCCTGAGA
3401 GAGGAGCAGG CCTTGAGCTG CCCAGGACAG AGAGCAGGAT GTCAGGGCCA
3451 TGGTGGGCCC AGGATTCCCC GGCTGGATTC CCCAGTGCTT AACTCTTCCT
3501 CCCTTCTCCA CAGCTTCCTA ACCAAGAGAG GCCGGCAGGT CTGTGCTGAC
3551 CCCAGTGAGG AGTGGGTCCA GAAATACGTC AGTGACCTGG AGCTGAGTGC

3601 CTGAGGGGTC CAGAAGCTTC GAGGCCCAGC GACCTCAGTG GGCCCAGTGG
3651 GGAGGAGCAG GAGCCTGAGC CTTGGGAACA TGCGTGTGAC CTCCACAGCT
3701 ACCTCTTCTA TGGACTGGTT ATTGCCAAAC AGCCACACTG TGGGACTCTT
3751 CTTAACTTAA ATTTTAATTT ATTTATACTA TTTAGTTTTT ATAATTTATT
3801 TTTGATTTCA CAGTGTGTTT GTGATTGTTT GCTCTGAGAG TTCCCCCTGT
3851 CCCCTCCACC TTCCCTCACA GTGTGTCTGG TGACAACCGA GTGGCTGTCA
3901 TCGGCCTGTG TAGGCAGTCA TGGCACCAAA GCCACCAGAC TGACAAATGT
3951 GTATCAGATG CTTTTGTTCA GGGCTGTGAT CGGCCTGGGC AAATAATAAA
4001 GATGTTCTTT TAAACGGTAA ACCAGTATTG AGTTTGGTTT TGTTTTTCTG
4051 GCAAATCAAA ATCACTGGTT AAGAGGAATC ATAGGCAAAG ATTAGGAAGA
4101 GGTGAAATGG AGGGAAATTG GGAGAGATGG GGAGCGCTGC GACAGAGTTA
4151 TCCACTTCAC AAAATTCTGG AACATTGAAA CTACGAATAT GTTATAACTC
4201 AAATCGTAAT ATGCACGCTC TAGGAGAATT AACTACTTGA ATGGCCACC
4251 TTAAGCAGAG TATTCTGTAG GGCATATTCA TGATGAATCA AGCTCTTAAT
4301 AGCAATTATT TACATTGTTG AGGCTTACTC CTCCTACTGA GTGCTTTTTA
4351 TACATTGTTC ATTTAATCTT ACCAATGCAA TAGTACAGCT TAGGTACTAT
4401 TAATACCTCC ACTTGACAGA AAAGTAACCC AGGGCTCAGA AAGGTTAGAC
4451 AACTTGGCTG AGGTTACACA GCACGTAAAC GGTCAATTGT GTTCCAAAAC
4501 TGGACTTTTA TTGAACTACA GACTATGCTG TTAACCATTG ACCAAGTTAT
4551 TTCCCAAAGT ATGACCCGCC TATACTCAAA TCTTACCCCA TTCTTTAACA
4601 GATGATACTT TATCCATTGC AACCACTTCC TGTCAGGATT CTGAGTTGAC
4651 ATAGAGTGTT TCAGCAGTGA TTATTTAAGC CAATTACATC AGGATCTTTA
4701 GGTGTAGACC TGGGAACTGA TATTTTTATC AAGCTCATGA GGTGTTCCAT
4751 AGCATGTTAA TGACTGAGAG CCACTGTCAA TAGAATTC

which corresponds to the cDNA sequence and derived amino acid sequence MIP-1α (macrophage inflammatory protein 1-α). See Blum et al., DNA Cell Biol. 9, 589-602 (1990).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 28) of:

Met Lys Leu Cys Val Thr Val Leu Ser Leu Leu Met Leu Val Ala Ala Phe Cys Ser Pro
Ala Leu Ser Ala Pro Met Gly Ser Asp Pro Pro Thr Ala Cys Cys Phe Ser Tyr Thr Ala
Arg Lys Leu Pro Arg Asn Phe Val Val Asp Tyr Tyr Glu Thr Ser Ser Leu Cys Ser Gln
Pro Ala Val Val Phe Gln Thr Lys Arg Ser Lys Gln Val Cys Ala Asp Pro Ser Glu Ser
Trp Val Gln Glu Tyr Val Tyr Asp Leu Glu Leu Asn

encoded by the cDNA sequence (SEQ ID NO: 29) of:

1 TTCCCCCCCC CCCCCCCCCC CCCCGCCCGA GCACAGGACA CAGCTGGGTT
51 CTGAAGCTTC TGAGTTCTGC AGCCTCACCT CTGAGAAAAC CTCTTTTCCA

101 CCAATACCAT GAAGCTCTGC GTGACTGTCC TGTCTCTCCT CATGCTAGTA
151 GCTGCCTTCT GCTCTCCAGC GCTCTCAGCA CCAATGGGCT CAGACCCTCC
201 CACCGCCTGC TGCTTTTCTT ACACCGCGAG GAAGCTTCCT CGCAACTTTG
251 TGGTAGATTA CTATGAGACC AGCAGCCTCT GCTCCCAGCC AGCTGTGGTA
301 TTCCAAACCA AAAGAAGCAA GCAAGTCTGT GCTGATCCCA GTGAATCCTG
351 GGTCCAGGAG TACGTGTATG ACCTGGAACT GAACTGAGCT GCTCAGAGAC
401 AGGAAGTCTT CAGGGAAGGT CACCTGAGCC CGGATGCTTC TCCATGAGAC
451 ACATCTCCTC CATACTCAGG ACTCCTCTCC GCAGTTCCTG TCCCTTCTCT
501 TAATTTAATC TTTTTTATGT GCCGTGTTAT TGTATTAGGT GTCATTTCCA
551 TTATTTATAT TAGTTTAGCC AAAGGATAAG TGTCCTATGG GGATGGTCCA
601 CTGTCACTGT TTCTCTGCTG TTGCAAATAC ATGGATAACA CATTTGATTC
651 TGTGTGTTTT CCATAATAAA ACTTTAAAAT AAAATGCAGA CAGTTA

which corresponds to the cDNA sequence and derived amino acid sequence MIP-1β (macrophage inflammatory protein 1-β). See Lipes et al., Proc. Natl. Acad. Sci. U.S.A. 85, 9704-9708 (1988).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 30) of:

Met Gln Ile Ile Thr Thr Ala Leu Val Cys Leu Leu Leu Ala Gly Met Trp Pro Glu Asp
Val Asp Ser Lys Ser Met Gln Val Pro Phe Ser Arg Cys Cys Phe Ser Phe Ala Glu Gln
Glu Ile Pro Leu Arg Ala Ile Leu Cys Tyr Arg Asn Thr Ser Ser Ile Cys Ser Asn Glu
Gly Leu Ile Phe Lys Leu Lys Arg Gly Lys Glu Ala Cys Ala Leu Asp Thr Val Gly Trp
Val Gln Arg His Arg Lys Met Leu Arg His Cys Pro Ser Lys Arg Lys

encoded by the cDNA sequence (SEQ ID NO: 31) of:

1 ACCAGGCTCA TCAAAGCTGC TCCAGGAAGG CCCAAGCCAG ACCAGAAGAC
51 ATGCAGATCA TCACCACAGC CCTGGTGTGC TTGCTGCTAG CTGGGATGTG
101 GCCGGAAGAT GTGGACAGCA AGAGCATGCA GGTACCCTTC TCCAGATGTT
151 GCTTCTCATT TGCGGAGCAA GAGATTCCCC TGAGGGCAAT CCTGTGTTAC
201 AGAAATACCA GCTCCATCTG CTCCAATGAG GGCTTAATAT TCAAGCTGAA
251 GAGAGGCAAA GAGGCCTGCG CCTTGGACAC AGTTGGATGG GTTCAGAGGC
301 ACAGAAAAAT GCTGAGGCAC TGCCCGTCAA AAAGAAAATG AGCAGATTTC
351 TTTCCATTGT GGGCTCTGGA AACCACATGG CTTCACCTGT CCCCGAAACT
401 ACCAGCCCTA CACCATTCCT TCTGCCCTGC TTTTGCTAGG TCACAGAGGA
451 TCTGCTTGGT CTTGATAAGC TATGTTGTTG CACTTTAAAC ATTTAAATTA
501 TACAATCATC AACCCCCAAC

which corresponds to the cDNA sequence and derived amino acid sequence human secreted protein (I-309). See Miller et al., J. Immunol. 143, 2907-2916 (1989).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 32) of:

Met Lys Val Ser Ala Ala Leu Leu Cys Leu Leu Leu Ile Ala Ala Thr Phe Ile Pro Gln
Gly Lys Ala Gln Pro Asp Ala Ile Asn Ala Pro Val Thr Cys Cys Tyr Asn Phe Thr Asn
Arg Lys Ile Ser Val Gln Arg Leu Ala Ser Tyr Arg Arg Ile Thr Ser Ser Lys Cys Pro Lys
Glu Ala Val Ile Phe Lys Thr Ile Val Ala Lys Glu Ile Cys Ala Asp Pro Lys Gln Lys Trp
Val Gln Asp Ser Met Asp His Leu Asp Lys Gln Thr Gln Thr Pro Lys Thr

encoded by the cDNA sequence (SEQ ID NO: 33) of:

```
   1 CTAACCCAGA AACATCCAAT TCTCAAACTG AAGCTCGCAC TCTCGCCTCC
  51 AGCATGAAAG TCTCTGCCGC CCTTCTGTGC CTGCTGCTCA TAGCAGCCAC
 101 CTTCATTCCC CAAGGGCTCG CTCAGCCAGA TGCAATCAAT GCCCCAGTCA
 151 CCTGCTGTTA TAACTTCACC AATAGGAAGA TCTCAGTGCA GAGGCTCGCG
 201 AGCTATAGAA GAATCACCAG CAGCAAGTGT CCCAAAGAAG CTGTGATCTT
 251 CAAGACCATT GTGGCCAAGG AGATCTGTGC TGACCCCAAG CAGAAGTGGG
 301 TTCAGGATTC CATGGACCAC CTGGACAAGC AAACCCAAAC TCCGAAGACT
 351 TGAACACTCA CTCCACAACC CAAGAATCTG CAGCTAACTT ATTTTCCCCT
 401 AGCTTTCCCC AGACACCCTG TTTTATTTTA TTATAATGAA TTTTGTTTGT
 451 TGATGTGAAA CATTATGCCT TAAGTAATGT TAATTCTTAT TTAAGTTATT
 501 GATGTTTTAA GTTTATCTTT CATGGTACTA GTGTTTTTTA GATACAGAGA
 551 CTTGGGGAAA TTGCTTTTCC TCTTGAACCA CAGTTCTACC CCTGGGATGT
 601 TTTGAGGGTC TTTGCAAGAA TCATTAATAC AAAGAATTTT TTTTAACATT
 651 CCAATGCATT GCTAAAATAT TATTGTGGAA ATGAATATTT TGTAACTATT
 701 ACACCAAATA AATATATTTT TGTAC
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which corresponds to the cDNA sequence and derived amino acid sequence monocyte chemoattractant protein 1 (MCP-1). See Yoshimura et al., FEBS Lett. 244, 487-493 (1989).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 34) of:

Met Lys Ala Ser Ala Ala Leu Leu Cys Leu Leu Leu Thr Ala Ala Ala Phe Ser Pro Gln
Gly Leu Ala Gln Pro Val Gly Ile Asn Thr Ser Thr Thr Cys Cys Tyr Arg Phe Ile Asn
Lys Lys Ile Pro Lys Gln Arg Leu Glu Ser Tyr Arg Arg Thr Thr Ser Ser His Cys Pro
Arg Glu Ala Val Ile Phe Lys Thr Lys Leu Asp Lys Glu Ile Cys Ala Asp Pro Thr Gln
Lys Trp Val Gln Asp Phe Met Lys His Leu Asp Lys Lys Thr Gln Thr Pro Lys Leu

encoded by the cDNA sequence (SEQ ID NO: 35) of:

```
   1 AGCAGAGGGG CTGAGACCAA ACCAGAAACC TCCAATTCTC ATGTGGAAGC
  51 CCATGCCCTC ACCCTCCAAC ATGAAAGCCT CTGCAGCACT TCTGTGTCTG
 101 CTGCTCACAG CAGCTGCTTT CAGCCCCCAG GGGCTTGCTC AGCCAGTTGG
```

151 GATTAATACT TCAACTACCT GCTGCTACAG ATTTATCAAT AAGAAAATCC
201 CTAAGCAGAG GCTGGAGAGC TACAGAAGGA CCACCAGTAG CCACTGTCCC
251 CGGGAAGCTG TAATCTTCAA GACCAAACTG GACAAGGAGA TCTGTGCTGA
301 CCCCACACAG AAGTGGGTCC AGGACTTTAT GAAGCACCTG GACAAGAAAA
351 CCCAAACTCC AAAGCTTTGA ACATTCATGA CTGAACTAAA AACAAGCCAT
401 GACTTGAGAA ACAAATAATT TGTATACCCT GTCCTTTCTC AGAGTGGTTC
451 TGAGATTATT TTAATCTAAT TCTAAGGAAT ATGAGCTTTA TGTAATAATG
501 TGAATCATGG TTTTTCTTAG TAGATTTTAA AAGTTATTAA TATTTTAATT
551 TAATCTTCCA TGGATTTTGG TGGGTTTTGA ACATAAAGCC TTGGATGTAT
601 ATGTCATCTC AGTGCTGTAA AAACTGTGGG ATGCTCCTCC CTTCTCTACC
651 TCATGGGGGT ATTGTATAAG TCCTTGCAAG AATCAGTGCA AAGATTTGCT
701 TTAATTGTTA AGATATGATG TCCCTATGGA AGCATATTGT TATTATATAA
751 TTACATATTT GCATATGTAT GACTCCCAAA TTTTCACATA AAATAGATTT
801 TTGTAAAAAA

which corresponds to the cDNA sequence and derived amino acid sequence monocyte chemoattractant protein 3 (MCP-3). See: Structural and Functional Identification of Two Human, Tumor-derived Monocyte Chemotactic Proteins (MCP-2 and MCP-3) Belonging to the Chemokine Family. Jo Van Damme, Paul Proost, Jean-Pierre Lenaerts, and Ghislain Opdenakker. J. Exp. Med. 176: 59-65, 1992.

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 36) of:

Met Lys Val Ser Ala Ala Arg Leu Ala Val Ile Leu Ile Ala Thr Ala Leu Cys Ala Pro Ala Ser Ala Ser Pro Tyr Ser Ser Asp Thr Thr Pro Cys Cys Phe Ala Tyr Ile Ala Arg Pro Leu Pro Arg Ala His Ile Lys Glu Tyr Phe Tyr Thr Ser Gly Lys Cys Ser Asn Pro Ala Val Val Phe Val Thr Arg Lys Asn Arg Gln Val Cys Ala Asn Pro Glu Lys Lys Trp Val Arg Glu Tyr Ile Asn Ser Leu Glu Met Ser

encoded by the cDNA sequence (SEQ ID NO: 37) of:

1 CCTCCGACAG CCTCTCCACA GGTACCATGA AGGTCTCCGC GGCACGCCTC
51 GCTGTCATCC TCATTGCTAC TGCCCTCTGC GCTCCTGCAT CTGCCTCCCC
101 ATATTCCTCG GACACCACAC CCTGCTGCTT TGCCTACATT GCCCGCCCAC
151 TGCCCCGTGC CCACATCAAG GAGTATTTCT ACACCAGTGG CAAGTGCTCC
201 AACCCAGCAG TCGTCTTTGT CACCCGAAAG AACCGCCAAG TGTGTGCCAA
251 CCCAGAGAAG AAATGGGTTC GGGAGTACAT CAACTCTTTG GAGATGAGCT
301 AGGATGGAGA GTCCTTGAAC CTGAACTTAC ACAAATTTGC CTGTTTCTGC
351 TTGCTCTTGT CCTAGCTTGG GAGGCTTCCC CTCACTATCC TACCCCACCC
401 GCTCCTTGAA GGGCCCAGAT TCTGACCACG ACGAGCAGCA GTTACAAAAA

451 CCTTCCCCAG GCTGGACGTG GTGGCTCAGC CTTGTAATCC CAGCACTTTG
501 GGAGGCCAAG GTGGGTGGAT CACTTGAGGT CAGGAGTTCG AGACAGCCTG
551 GCCAACATGA TGAAACCCCA TGTGTACTAA AAATACAAAA AATTAGCCGG
601 GCGTGGTAGC GGGCGCCTGT AGTCCCAGCT ACTCGGGAGG CTGAGGCAGG
651 AGAATGGCGT GAACCCGGGA GCGGAGCTTG CAGTGAGCCG AGATCGCGCC
701 ACTGCACTCC AGCCTGGGCG ACAGAGCGAG ACTCCGTCTC AAAAAAAAAA
751 AAAAAAAAAA AAAAAATACA AAAATTAGCC GCGTGGTGGC CCACGCCTGT
801 AATCCCAGCT ACTCGGGAGG CTAAGGCAGG AAAATTGTTT GAACCCAGGA
851 GGTGGAGGCT GCAGTGAGCT GAGATTGTGC CACTTCACTC CAGCCTGGGT
901 GACAAAGTGA GACTCCGTCA CAACAACAAC AACAAAAAGC TTCCCCAACT
951 AAAGCCTAGA AGAGCTTCTG AGGCGCTGCT TTGTCAAAAG GAAGTCTCTA
1001 GGTTCTGAGC TCTGGCTTTG CCTTGGCTTT GCAAGGGCTC TGTGACAAGG
1051 AAGGAAGTCA GCATGCCTCT AGAGGCAAGG AAGGGAGGAA CACTGCACTC
1101 TTAAGCTTCC GCCGTCTCAA CCCCTCACAG GAGCTTACTG GCAAACATGA
1151 AAAATCGGGG

which corresponds to the cDNA sequence and derived amino acid sequence Human T cell-specific protein (RANTES). See Schall et al., J. Immunol. 141, 1018-1025 (1988).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 38) of:

Met Arg Ile Ser Ala Thr Leu Leu Cys Leu Leu Leu Ile Ala Ala Ala Phe Ser Ile Gln Val Trp Ala Gln Pro Asp Gly Pro Asn Ala Ser Thr Cys Cys Tyr Val Lys Lys Gln Lys Ile Pro Lys Arg Asn Leu Lys Ser Tyr Arg Arg Ile Thr Ser Ser Arg Cys Pro Trp Glu Ala Val Ile Phe Lys Thr Lys Lys Gly Met Glu Val Cys Arg Glu Ala His Gln Lys Trp Val Glu Glu Ala Ile Ala Tyr Leu Asp Met Lys Thr Pro Thr Pro Lys Pro

encoded by the cDNA sequence (SEQ ID NO: 39) of:

1 ACTGAAGCCA GCTCTCTCAC TCTCTTTCTC CACCATGAGG ATCTCTGCCA
51 CGCTTCTGTG CCTGCTGCTC ATAGCCGCTG CTTTCAGCAT CCAAGTGTGG
101 GCCCAACCAG ATGGGCCCAA TGCATCCACA TGCTGCTATG TCAAGAAACA
151 AAAGATCCCC AAGAGGAATC TCAAGAGCTA CAGAAGGATC ACCAGTAGTC
201 GGTGTCCCTG GGAAGCTGTT ATCTTCAAGA CAAAGAAGGG CATGGAAGTC
251 TGTCGTGAAG CCCATCAGAA GTGGGTCGAG GAGGCTATAG CATACTTAGA
301 CATGAAAACC CCAACTCCAA AGCCTTGAAG AAATGTGCCT GAACAGAAAC
351 CAACCTAGGA GCCAAGAAGC AAAAATTCCT CACCGCTGTT CTTTCTGAGA
401 ACTGTTGATG AAATGTGTTG ATCACGGTCC TAAGGGATAG GAGCTGTCTG
451 TAGGAATGTG AAACAGTCAC GCCTAAGGAA TGGTCTTTAA GTTATTAATA
501 TTTTTATTTA ATTAGCCATG TACTTTGGTG TGATTTGAAT GTAAAGCTCT

551 GGAGACCTCA TGTCACTTTA ACATTGTGTT AGCTGCAGAA TTC

which corresponds to the cDNA sequence and derived amino acid sequence human *fic* (growth factor-activated gene). See Heinrich et al., Molecular and Cellular Biology 13: 2020-2030, 1993.

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 40) of:

Asp Ser Val Ser Ile Phe Ile Thr Cys Cys Phe Asn Val Ile Asn Arg Lys Ile Pro Ile Gln Arg Leu Glu Ser Tyr Thr Arg Ile Thr Asn Ile Gln Cys Pro Lys Glu Ala Val Ile Phe Lys Thr Gly Lys Glu Val Cys Ala Asp Pro Lys Glu Arg Trp Val Arg Asp Ser Met Lys His Lys Asp Gln Ile Phe Gln Asn Leu Lys Pro

which corresponds to the cDNA sequence and derived amino acid sequence monocyte chemoattractant protein 2 (MCP-2). See VanDamme et al., J. Exp. Med. 176: 59 - 65, 1992.

The purified heparanase of the present invention, allows for the convenient selection of compounds having anti-heparanase activity (AHA compounds), i.e. inhibitors of heparanase activity (IHA), by measuring inhibition of heparanase activity. Inhibition of heparanase activity can be measured utilizing *in vivo* radiolabeled heparan sulfate/heparin. This ligand is radiolabeled to high specific activity by intraperitoneal injection of 0.5mCi of S-35 sulfate into C57 mice bearing a 1-2 cm basement membrane tumor (EHS; Engelbreth, Holm, Swarm tumor). The tumor is harvested after 16 hours and the heparan sulfate proteoglycan extracted in 4 volumes of 6M urea, 20mM Tris pH 6.8, protease inhibitors, 0.15M NaCl and 0.5% triton X-100. The urea extract is chromatographed on an anion exchange column and the proteoglycan is eluted in a linear gradient of NaCl. The radiolabeled proteoglycan is exchanged into a solution of 4.0M guanidine-HCl, 20mM Tris pH 7.4 and applied to a size exclusion column. The proteoglycan peak is pooled and exchanged into 0.15mM NaCl and 20mM Tris pH7.4.

Purified, radiolabeled proteoglycan is coupled to commercially available agarose support. A quantitative assay of heparanase activity is constructed with the radiolabeled ligand in a multi-well format. Briefly, known quantities of recombinant heparanase are added to a multi-well plate containing equal amounts of radiolabeled ligand in each well. Enzyme-ligand interaction proceeds overnight and the ligand-agarose complex is recovered by centrifugation. Radioactivity in the liquid phase is determined by scintillation counting and is the measure of enzyme activity. Potential enzyme inhibitors can be evaluated by adding the compound to the solution phase or alternatively adding the assay components to multi-well plates containing preweighed amounts of test compound.

In addition, the purified heparanase of the subject invention can be used for therapeutic wound healing or can be immobilized onto filters and used to degrade heparin from the blood of patients post-surgery.

Wound treatment can be achieved by administration to an afflicted individual an effective amount of a pharmaceutical composition comprising the purified heparanase in combination with a pharmaceutically acceptable, preferably slow releasing, carrier. See. e.g. PCT/US90/04772, incorporated herein by reference.

Immobilization onto filters can be achieved by the methods well known in the art including those disclosed by Langer et al. in *Biomaterials: Inter-facial Phenomenon and Applications*, eds. Cooper et al, pp 493-509, 1982 and those described in U.S. Patent No. 4,373,023, 4,863,611 and 5,211,850 (all incorporated herein by reference).

The purified heparanase of the subject invention can be prepared by the method described in procedure A or procedure B, but preferably procedure A.

PROCEDURE A

Reverse transcription of the mRNA from activated human leukocyte-derived cells [preferably lymphocytes, neutrophils, platelets, Jurkatt lymphoma cells, Dami cells (Greenberg et al., Blood 72:1968-1977, (1988)] is used to prepare the cDNA for the desired heparanase enzyme (preferably SEQ. ID. NO: 1; optionally SEQ. ID. NO: 3, SEQ. ID. NO: 5, SEQ. ID. NO: 7; SEQ. ID. NO: 13, SEQ. ID. NO: 15, SEQ. ID. NO: 17, SEQ. ID. NO: 19, SEQ. ID. NO: 21, SEQ. ID. NO: 23, SEQ. ID. NO: 25, SEQ. ID. NO: 27, SEQ. ID. NO: 29; SEQ. ID. NO: 31, SEQ. ID. NO: 33 or SEQ. ID. NO: 35), employing standard PCR cloning techniques (described in Sambrook et al., in: *Molecular Cloning, A Laboratory Manual.* Second Edition, 1989. Cold Spring Harbor Press). The cDNA encoding the heparanase enzyme is cloned into XbaI/BamH1 sites in the commercially available baculovirus vector pVL 1392 (Pharmingen; San Diego, CA). High titer infectious virus is selected for use in infecting sf9 insect cells (Luckow and Summers, *Bio/Technology*. 6,47 1988). Serum-free medium conditioned by infected sf9 cells is collected after 72 hours. This media is the starting material for purification of recombinant heparanase. Serum-free conditioned media is adjusted to contain 20mM Sodium Acetate, pH 5.0, 0.15M NaCl, 1mM reduced glutathione (GSH), 1mM dithiothreitol (DTT) and 10mM beta-octylglucoside. Medium is applied to a column of cation-exchange resin (Pharmacia) and eluted from the column in a linear gradient of NaCl. Fractions containing heparanase are pooled and diluted to a final salt concentration of 0.15M NaCl. To this solution is added 20mM Tris and the pH adjusted to 7.0. The solution is applied to a column of heparin-Sepharose (Pharmacia) and eluted with a linear salt gradient buffered to pH 5.0 with 20mM Sodium Acetate. Heparanase is concentrated to 0.5mg/ml in an Amicon concentrator fitted with a YM-2 membrane and stored at -80 degrees. For optimal activity (greater than 50 units heparanase activity per ug protein) incubation in the presence of transglutaminase, under reducing conditions, in accordance with the procedure in Example 2, Part C.

PROCEDURE B

This procedure describes the purification to homogeneity of heparanase (SEQ. ID. NO: 1) from human blood cells or cell lines (such as platelets) under reducing conditions which allow for the occurrence of post-translational modifications that increase the specific activity of heparanase and make it suitable for use in the above described screening assay. The cells are treated with a suitable activator (such as, but not limited to, thrombin or histamine) which allows for the release of enzymes and cytokines from the cell. Reducing agents are added to the supernatant from the activated cells. Suitable reducing agents include dithiothreitol (DTT), dithioerythritol (DTE), reduced glutathione (GSH), and β-mercaptoethanol. The reduced, activated supernatant is chromatographed on a column of immobilized heparin or heparan sulfate under reducing conditions at pH 5, using a salt gradient (such as NaCl, KCl, or other salt) to elute the bound proteins. Fractions containing heparanase activity are pooled and exchanged into any buffer appropriate for the pH of 6.8 and containing 0.15 M NaCl, reducing agents, and non-ionic detergent. This is passed over any suitable anion-exchange column (bed volume of 5 ml or less). The unbound material from this column is adjusted to pH 5 with acid, and is loaded onto any suitable cation-exchange column (bed volume of 5 ml or less), equilibrated in a suitable pH 5 buffer containing 0.15 M NaCl, reducing agents, and non-ionic detergents. The bound protein is eluted from the column with a salt gradient, and the fractions containing heparanase activity are pooled and size fractionated to below 30,000 daltons with 30 K-cut-off membranes. The protein below 30,000 daltons is concentrated by either heparin-sepharose chromatography or by centrifugation through 5 K-cut-off membranes.

The present invention is seen more fully by the examples set forth below.

Example 1: Use of Heparanase as a screen for AHA compounds.

1. Heparan sulfate, metabolically labeled (S-35) to a high-specific activity- as described above for the EHS tumor, prepared by papain digestion of chromatographically purified heparan sulfate proteoglycan is coupled to cyanogen bromide activated Sepharose-6B (Pharmacia) according to manufacturer's instructions.

2. ^{35}S-Heparan sulfate-Sepharose 6B is resuspended in: 0.15 M NaCl, 0.03% human serum albumin, 10 μM $MgCl_2$, 10 μM $CaCl_2$, antiproteolytic agents (1 μg/ml leupeptin, 2 μg/ml antipain, 10 μg/ml benzamidine, 10 units/ml aprotinin, 1 μg/ml chymostatin, and 1 μg/ml pepstatin), and 0.05 M Na acetate, pH 5.6 and 5,000 cpm, in a total volume of 200 μl, are aliquoted into each well of a 96 well plate. To each well is added 5 units of activated heparanase and the digestion allowed to proceed overnight at 37 degrees.

3. Separation of digested product is accomplished by centrifugation of the 96 well plate. The supernatant, containing cleaved heparan sulfate is decanted and quantitated by scintillation counting.

4. Inhibitors of heparanase activity can be introduced into the liquid-phase of the assay.

5. A potential inhibitor of heparanase activity would be identified by its ability to reduce the amount of radiolabeled heparan sulfate released into the supernatant by 50% at a concentration of 1 μM or less.

Example 2: The preparation of heparanase under reducing conditions as outlined in Procedure B.

Part A:

Platelet-rich plasma (10^9 platelets/ml; 1800 ml) is obtained from healthy, informed volunteers by plasmapheresis. The plasma is removed from the platelets by centrifugation (Heldin, *et al., Exp. Cell Res.* 109: 429-437, 1977). Platelets suspended in phosphate buffered saline (PBS; 0.1 original volume) are then stimulated with 1 U/ml thrombin for 5 min at 37°C. This concentration of thrombin was reported to release 100% of the heparanase activity from platelets (Oldberg, *et al., Biochemistry* **19**: 5755-5762, 1980). Following activation, the thrombin is inactivated by the addition of 100 mM phenylmethylsulfonylfluoride (PMSF), and the platelets are centrifuged at 2000 x g for 30 min at 4°C. The supernatant is stored at -80°C until used for the chromatographic purification of heparanase (Part B).

Part B: Chromatographic purification of heparanase.

1. Heparin-Sepharose Chromatography. Activated platelet supernatants are pooled and adjusted to contain 1 mM GSH and 1 mM DTT. This pool is loaded (2.5 ml/min) onto a column of heparin-sepharose (2.6 x 7.5 cm, 40 ml) equilibrated in 1 mM GSH, 1 mM DTT, 150 mM NaCl, 10 mM $NaPO_4$, pH 7.4. After loading the sample, the column is washed with 200 ml of 0.15 M NaCl, 1 mM GSH, 1 mM DTT, 10 mM Na acetate, pH 5, followed by 60 ml of 0.35 M NaCl, 1 mM DTT, 1 mM GSH, 10 mM Na acetate, pH 5. The column is then eluted with a 160 ml linear gradient between 0.35 M NaCl and 1.5 M NaCl in the same buffer. Aliquots of each fraction are used for determination of heparanase activity by the "Purification Assay" described later.

2. Anion-exchange chromatography (For example, DEAE-Sephacel, Pharmacia). The 0.9 M - 1.15 M NaCl fractions from the heparin-sepharose column are concentrated using a stirred cell fitted with a PM-10 membrane, and the buffer is exchanged to 0.15 M NaCl, 1 mM DTT, 1 mM GSH, 10 mM β-octylglucoside, 10 mM sodium phosphate, pH 6.8 (8 ml). This sample is loaded onto a 5 ml column of DEAE-Sephacel equilibrated in the same buffer. After loading, the column is washed to baseline absorbance (280 nm) with the equilibration buffer. The flow-through and wash with equilibration buffer are collected as one pool. The column is then eluted with 10 ml of 0.15 M NaCl, 10 mM β-octylglucoside, 1 mM GSH, 1 mM DTT, 10 mM Na acetate, pH 5, followed by 10 ml of 1.5 M NaCl, 10 mM β-octylglucoside, 1 mM GSH, 1 mM

DTT, 10 mM Na acetate, pH 5. Aliquots of each pool are used for determination of heparanase activity by the "Purification Assay".

3. Cation Exchange. The unbound sample from the DEAE-Sephacel column is adjusted to pH 5 with glacial acetic acid and loaded onto a cation exchange column (Poros HS/F, 4.6 mm x 50 mm; PerSeptive Biosystems), pre-equilibrated with 0.15 M NaCl, 1 mM DTT, 1 mM GSH, 10 mM β-octylglucoside, 10 mM Na acetate, pH 5. The flow rate is 3.0 ml/min and 3 ml fractions are collected. After washing the column with 35 ml of equilibration buffer, the column is developed with a 55 ml linear gradient between 0.15 M and 1.5 M NaCl in the same buffer. 10 µl aliquots of the fractions are used for determination of heparanase activity by the "Purification Assay".

4. Size fractionation to < 30 kD and concentration on immobilized heparin (Hi-trap heparin-sepharose, Pharmacia). The activity from the Poros HS/F column is size fractionated by centrifuging through 30,000 molecular weight cut-off filters (Millipore ultrafree-MC 30,000 NMWL filter units). The < 30 kD pool is diluted to contain 0.15 M NaCl, and is loaded onto a 1 ml Hi-trap heparin column, pre-equilibrated with 0.15 M NaCl, 1 mM DTT, 1 mM GSH, 10 mM Na acetate, pH 5. The column is eluted with 1.2 M NaCl in the same buffer and the single eluted peak contains the heparanase activity.

Part C: Properties of the purified heparanase.

The final yield of heparanase protein from 1850 ml platelet-rich plasma was 2.7 mg. Protein concentration was determined by the method of Lowry (*J. Biol. Chem.* **193:** 265-275, 1951), or if more precise determinations were required, by amino acid analysis on an amino acid analyzer (Beckman 6300). The overall recovery of activity was 8%, with a 4150-fold purification. The preparation was judged to be homogeneous by the presence of a single band of 9000 daltons on an 18% silver-stained SDS-polyacrylamide gel, run according to the method of Laemmli (*Nature* **227:** 680-685, 1970).

The pH optimum of the purified heparanase was determined by conducting the "Purification assay" activity between pH 3.5 and 8.0, using a citrate buffer (pH 3.5 - 6.0), citrate-phosphate buffer (pH 6.5 - 7.0), and phosphate buffer (pH 7.5 - 8.). Heparanase was active between pH 5.0 and 8.0, with the optimum pH at 5.8.

N-terminal amino acid sequencing of heparanase produced by this procedure was performed using a gas/liquid phase Protein Sequencer (Applied Biosystems Inc. Model 470). Phenylthiodantoin amino acids were resolved and quantitated by an on-line HPLC system (Model 120, Applied Biosystems Inc.) with data analysis on a Nelson Analytical System. N-terminal amino acid sequences of the heparanase produced in this example were 85 % SEQ. ID. NO: 9 (namely:

Asn Leu Ala Lys Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Tyr Ala
1 5 10 15

Glu Leu Arg),

which is identical to CTAP-III, and 15% SEQ. ID. NO: 10 (namely:

Ser Ser Thr Lys Gly Gln Thr Lys Arg Asn Leu Ala Lys Gly Lys Glu),
1 5 10 15

which is the precursor form, platelet basic protein. Interestingly, the N-terminal sequence of commercial β-thromboglobulin (namely, Calbiochem (Cat. # 605165), Celsus Laboratories (Cat. # 41705), and Haematologic Technologies (Cat. # HBTG-0210), which has low levels of heparanase activity, was 100% SEQ. ID. NO: 11 (namely:

Asn Leu Ala Lys Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Tyr Ala Glu),
1 5 10 15

indicating that the commercial preparation is actually CTAP-III and not β-thromboglobulin.

Chromatofocusing of the heparanase produced by this procedure results in two peaks of differing isoelectric points. To perform the chromatofocusing, heparanase is dissolved in 0.025 M imidazole, pH 7.3. The sample is loaded onto a 0.5 x 20 cm column of Polybuffer Exchanger 94 (Pharmacia), equilibrated with 0.025 M imidazole, pH 7.3. Immediately after sample loading, Polybuffer 74 (Pharmacia; 1:8, pH 4) is pumped onto the column at 0.5 ml/min. 2 ml fractions are collected, and the pH of each fraction is determined by a narrow range (pH 4-7) pH paper. Aliquots of each fraction are used to determine heparanase activity by the "Purification Assay." All of the activity is associated with an absorbance (280 nm) peak that eluted at pH 4.8 to 5.1, representing approximately 10% of total protein, while 90% of the protein is eluted at pH 7.3 and is inactive. Aliquots of each protein peak are separated from the ampholytes by C_4 reverse phase chromatography. The peak that eluted from the chromatofocusing column at pH 7.3 has *N*-terminal sequences for platelet basic protein and the processed form, CTAP-III. The peak that is eluted from the chromatofocusing column at pH 4.8 - 5.1 also contains the sequences of platelet basic protein and the processed form, CTAP-III. All of the platelet basic protein processed forms have pI's that are calculated and reported to be greater than 7.6. Thus, the heparanase activity resides in the platelet basic protein and/or the processed form, CTAP-III that is modified such that the pI is lowered to 4.8 - 5.1.

Heparanase obtained after chromatofocusing exhibited a specific activity of 80 units/μg protein, using the "Purification Assay." This represents a 1000-fold increase in the specific activity compared to the commercial protein (β-thromboglobulin; 0.075 units/μg protein).

The modification that may be responsible for the lower isoelectric point of active heparanase is ADP-ribosylation. ADP-ribosylation (Adenine diphosphate-ribosylation) is a post-translational modification of proteins or DNA in which the ADP-ribose group of NAD (Nicotinamide adenine dinucleotide) is enzymatically transferred to proteins or DNA. Since this

modification adds two negatively charged phosphate groups to a molecule, it would result in a lower isoelectric point. Activated platelet supernatants were incubated in 1 mM DTT, 2 mM $MgCl_2$, 100 mM HEPES, pH 7.4, and 0.5 μM [^{32}P]NAD (Specific activity = 1000 Ci/mmol). The labeled proteins were separated by SDS-polyacrylamide gel electrophoresis on an 18% gel, transferred to PVDF (polyvinylidene difluoride) membrane, and exposed to X-ray film. The autoradiogram demonstrates the incorporation of [^{32}P] into a protein of 8000 daltons. The PVDF membrane was immunoblotted with the anti-Peptide C antisera (1:1500 in PBS containing 5% dry milk, 0.05% Tween-20, 0.15 M NaCl, 20 mM Tris, pH 7.4, 2 hours at room temperature, followed by incubation with peroxidase-labeled goat anti-chicken IgG (1:500 in above buffer, 1 hour room temperature), and reacted with a peroxidase substrate. The immunoblot revealed that the 8000 dalton that was labeled with [^{32}P] was CTAP-III/heparanase. The addition of 200 μM sodium nitroprusside, a spontaneous releaser of nitric oxide, to the ADP-ribosylation reaction resulted in 5-fold more incorporation of [^{32}P] label into CTAP-III/heparanase, suggesting that this modification can be regulated in vivo by nitric oxide. Finally, in an analgous manner to that of glyceraldehyde-3-phosphate dehydrogenase, another platelet ADP-ribosylated glycolytic enzyme (Zhang and Snyder, *Proc. Natl. Acad. Sci. USA* 89: 9382-9385), it was determined that CTAP-III/heparanase has an auto-ADP-ribosylation activity, since the [^{32}P]-ADP-ribosylation of CTAP-III/heparanase occurs in reactions where the only protein present is commercial CTAP-III or purified heparanase. Other chemokine family members tested, which includes IP-1-, IL-8, gro-α, and MCAF, also have auto-ADP-ribosylation activity.

It is contemplated that the high specific activity of CTAP-III/heparanase is a consequence of ADP-ribosylation of the enzyme in the presence of nitric oxide. It is further contemplated that the action of transglutaminase on the ADP-ribosylated enzyme will lead to further increase in the specific activity.

An amino acid composition of the heparanase produced in Example 2 gave the expected amino acid composition for CTAP-III and N-terminal sequencing revealed sequences for platelet basic protein and the processed form, CTAP-III , confirming that the heparanase activity is contained in this set of processed proteins and is not due to a minor contaminant. The presence of heparanase activity in three commercial sources of β-thromboglobulin also confirms this conclusion. In addition, polyclonal antibodies to β-thromboglobulin were found to precipitate 30 - 70% of the heparanase activity in three separate experiments, providing additional confirmation.

The activation of heparanase with transglutaminase (prepared in accordance with Example 2, Part B) results in a substantial (about 13-fold) increase in the specific activity of the enzyme. The heparanase (2 *ul* at 56 nM) obtained by Example 2, Part B is treated with either

transglutaminase from guinea pig liver (4 mU; Sigma) or with Factor XIII (1 µg; Celsus Laboratories, Inc.), the blood coagulation factor that is activated by treatment with 5 units of thrombin at 37 degrees for 30 minutes. Heparanase is activated by incubation of either 2mU liver transglutaminase or 5 units of activated Factor XIII in the presence of 0.1M NaAcetate buffer at pH 6.0 containing 1mM reduced glutathione and 1mM CaCl for 35 minutes at 37 degrees. Treatment of heparanase with either type of transglutaminase results in a substantial increase in the specific activity of the heparanase.

The high degree of sequence identity between CTAP-III and Interleukin-8, a CXC chemokine family member, assures that an essentially identical folding pattern will be shared by the two proteins. Since the 3-dimensional structure of Interleukin-8 is known (Clore, *et al.*, *Biochemistry* **29**: 1689-1696,1990; Baldwin, *et al., J. Biol. Chem.* **265**: 6851-6853), one can model the same for CTAP-III. Such a model can serve to direct research into rationally designed IHA and to help explain the action of transglutaminase in activating the CTAP-III.

Part D: Purification Assay for Heparanase Activity

Heparanase activity from platelets or column fractions is detected by its ability to digest the ≥ 70 kD ^{35}S-HSPG to produce lower molecular weight products. Each digest contains 10 µl sample, ^{35}S-HSPG (2000 cpm), 0.15 M NaCl, 0.03% human serum albumin, 10 µM $MgCl_2$, 10 µM $CaCl_2$, antiproteolytic agents (1 µg/ml leupeptin, 2 µg/ml antipain, 10 µg/ml benzamidine, 10 units/ml aprotinin, 1 µg/ml chymostatin, and 1 µg/ml pepstatin), and 0.05 M Na acetate, pH 5.6 in a total volume of 300 µl. Digests are carried out for 3 to 21 h. The presence of lower molecular weight radiolabeled products is detected by centrifugation through 30,000 MW-cutoff filters. The digests containing 2000 cpm of ^{35}S-HSPG (> 70 K) are centrifuged through 30,000 molecular weight cut-off filters (Millipore Ultrafree-MC 30,000 NMWL filter units). ^{35}S-HSPG degradation is evident by the presence of radioactivity in the filtrate that passed through the 30 K membrane; this heparanase activity is expressed as the % of total cpm < 30 K for a given digest. Analysis of heparan sulfate degradation by this method is quick and reproducible. 1 unit of heparanase activity is defined as 1% cpm < 30 K per h. For pH optimum determination, the 0.1 M Na acetate buffer is replaced by 50 mM citrate, citrate-phosphate, or phosphate buffer at varying pH's. For samples from chromatographic steps performed under reducing conditions (1 mM GSH, 1 mM DTT), the concentration of a thiol oxidant (diamide) needed for optimum activity is determined. This concentration (100 µM diamide) is added to all assay tubes when reduced samples are assayed.

Preparation of ^{35}S-HSPG (>70 K) for use in the "Purification Assay."

^{35}S-HSPG (>70 K) is prepared from mice bearing a basement membrane tumor that overproduces HSPG (EHS tumor), using modifications of the method of Ledbetter, et. al., 1987. Briefly, the radiolabeled HSPG was prepared by injecting C57BL mice bearing the EHS tumor

(Orkin, et.al., 1977) with sodium [^{35}S]sulfate (0.5 mCi/mouse) 18 h before harvesting the tumor. The HSPG is extracted from the weighed tumor with 6 volumes (w/v) of Buffer A (3.4 M NaCl, 0.1 M 6-aminohexanoic acid, 0.04 M EDTA, 0.008 M *N*-ethylmaleimide, 0.002 M PMSF, and 0.05 M Tris-HCl, pH 6.8), by homogenization with a Polytron for 30 s, followed by stirring at 4°C for 1 h. Insoluble material is collected by centrifugation (12,000 x g for 10 min), and the supernatant is discarded. The insoluble residue is reextracted with 2 volumes (original tumor weight) of Buffer A for 30 min with stirring at 4°C. Insoluble material is again collected by centrifugation, and the supernatant fraction is discarded. The insoluble material is then suspended in 6 volumes of Buffer B (6 M urea, 0.1 M 6-aminohexanoic acid, 0.04 M ethylenediaminetetraacetic acid (EDTA), 0.002 M PMSF, and 0.05 M Tris-HCl, pH 6.8), homogenized with an electric homogenizer (Polytron) for 30 s, and stirred for 2 h at 4°C. The mixture is centrifuged to remove insoluble material, and the supernatant is retained. The insoluble material is reextracted with 2 volumes of Buffer B. The mixture is centrifuged, and the supernatant is combined with the previous supernatant.

^{35}S-HSPG is isolated from the Buffer B supernatant by sequential chromatography on anion exchange and gel filtration columns. The Buffer B supernatant is dialyzed overnight against 10 volumes of 6 M urea, 0.15 M NaCl, 0.05 M Tris-HCl, pH 6.8, and is adjusted to contain 0.5% non-ionic detergent (Triton X-100). This supernatant (from 11 g tumor) is chromatographed on a 30 ml column of anion exchange resin (DEAE-Sephacel) equilibrated with 6 M urea, 0.15 M NaCl, 0.05% Triton X-100, 0.05 M Tris-HCl, pH 6.8. After loading the supernatant and washing with the equilibration buffer, the column is developed with a 250 ml linear gradient between 0.15 M NaCl and 1.15 M NaCl (Flow = 2.0 ml/min). Fractions are sampled for radioactivity, and those containing the $^{35}SO_4$ label that elutes from the DEAE-Sephacel between 0.4 M and 0.8 M NaCl are pooled. The proteoglycan is precipitated by the addition of 4 volumes of 100% EtOH at -20°C overnight. The precipitate is collected by centrifugation and is solubilized in 1 ml of Buffer C (4 M Gu-HCl, 20 mM Tris-HCl, pH 7.2). This solubilized pellet is used for chromatography on a calibrated gel filtration column (1.0 x 50 cm column of Superose 6; Pharmacia) equilibrated in Buffer C (Flow = 0.5 ml/min). Fractions are sampled for radioactivity, and those containing the $^{35}SO_4$ label that elutes with a molecular weight ≥ 70 kD were pooled. The proteoglycan is precipitated with 100% EtOH as described above. The pellet is dissolved in 3 ml PBS, and dialyzed against 3 x 100 volumes of PBS. Each preparation of ^{35}S-HSPG is confirmed to be ≥ 98% heparan sulfate by susceptibility to low pH nitrous acid degradation (Shiveley and Conrad, *Biochemistry* **15**: 3932-3942, 1976).

Example 3: Preparation of cDNA encoding Heparanase.

Media is removed from cultured HEL (HEL 92.1.7; Human erythroleukemia; ATCC No. TIB 180) cells stimulated with 10nM phorbol 12-myristate 13-acetate (Sigma Chemical Co., St.

Louis, MO) and the cells scraped from the dish and pelleted by centrifugation. The pellet is extracted with 200*u*l of TRI reagent (Molecular Research Center Inc. Cincinnati, OH) and the total cellular RNA is prepared according to the manufacturer's instructions. To prepare first strand synthesis the reverse transcriptase reaction was performed with 10*u*l of total cellular RNA in the presence of 4*u*l of 5x transcriptase buffer (Bethesda Research Laboratories, Gaithersburg, MD), 1*u*l 0.2mM DTT, 4*u*l random hexanucleotides (Amersham Corp. Arlington Heights, ILL), and 1*u*l 10mM dNTP (BRL). This solution is heated to 95 degrees C for 5 minutes and then placed on ice. To this is added 1*u*l RNAsin and 1*u*l reverse transcriptase (M-MLV), (Promega, Madison WI). This is incubated at 37 degrees for 60 minutes and then placed on ice. The polymerase chain reaction is carried out as follows. To 3*u*l of the first strand (above) is added 1*u*l of each Primer (see below), 77*u*l of water 10*u*l 10x PCR buffer (Perkin Elmer Cetus, Norwalk CT) and 2*u*l each dNTP. This solution is heat denatured at 95 degrees C and 1*u*l Amplitaq DNA polymerase (Perkin Elmer Cetus) is added. Hybridization temperature begins at 72 degrees and is lowered by one degree per cycle until reaching 55 degrees. Each hybridization step is followed with a constant elongation temperature of 72 degrees. Upon completion the solution is left at 0 degrees until storage at -20 degrees. The products of the PCR reaction are electrophoresed on 3% NuSieve, 1% agarose gels and bands of expected size are excised and purified by standard procedures.

Primers:

Platelet Basic Protein: TGG ACT AGT ATG TCC TCC ACC AAA GGA CAA ACT AA

CTAP III: TGG ACT AGT ATG AAC TTG GCG AAA GAG GA

B-thrombglobulin: TGG ACT AGT ATG GGC AAA GAG GAA AGT CTA GAC AG

NAP-2: TGG ACT AGT ATG GAA CTC CGC TGC ATG TGT ATA AA

Example 4: Preparation of cDNA encoding Heparanase.

Media is removed from cultured leukocyte-derived cells [e.g., lymphocytes, neutrophils, platelets, Jurkatt lymphoma cells, Dami cells (Greenberg et al., Blood 72:1968-1977, (1988)], stimulated with Concanavalin A or phorbol 12-myristate 13 acetate (Sigma Chemical Co., St. Louis, MO) and the cells scraped from the dish and pelleted by centrifugation. The pellet is extracted with 200*u*l of TRI reagent (Molecular Research Center Inc. Cincinnati, OH) and the total cellular RNA is prepared according to the manufacturer's instructions. To prepare first strand synthesis the reverse transcriptase reaction was performed with 10*u*l of total cellular RNA in the presence of 4*u*l of 5x transcriptase buffer (Bethesda Research Laboratories, Gaithersburg, MD), 1*u*l 0.2mM DTT, 4*u*l random hexanucleotides (Amersham Corp. Arlington Heights, ILL), and 1*u*l 10mM dNTP (BRL). This solution is heated to 95 degrees C for 5 minutes and then placed on ice. To this is added 1*u*l RNAsin and 1*u*l reverse transcriptase (M-MLV), (Promega,

Madison WI). This is incubated at 37 degrees for 60 minutes and then placed on ice. The polymerase chain reaction is carried out as follows. To 3*ul* of the first strand (above) is added 1*ul* of each Primer (see below), 77*ul* of water 10*ul* 10x PCR buffer (Perkin Elmer Cetus, Norwalk CT) and 2*ul* each dNTP. This solution is heat denatured at 95 degrees C and 1*ul* Amplitaq DNA polymerase (Perkin Elmer Cetus) is added. Hybridization temperature begins at 72 degrees and is lowered by one degree per cycle until reaching 55 degrees. Each hybridization step is followed with a constant elongation temperature of 72 degrees. Upon completion the solution is left at 0 degrees until storage at -20 degrees. The products of the PCR reaction are electrophoresed on 3% NuSieve, 1% agarose gels and bands of expected size are excised and purified by standard procedures.

Primers:

Platelet Basic Protein: TGG ACT AGT ATG TCC TCC ACC AAA GGA CAA ACT AA

CTAP III: TGG ACT AGT ATG AAC TTG GCG AAA GAG GA

B-thrombglobulin: TGG ACT AGT ATG GGC AAA GAG GAA AGT CTA GAC AG

NAP-2: TGG ACT AGT ATG GAA CTC CGC TGC ATG TGT ATA AA

All temperatures expressed throughout the subject specification are in degrees Centigrade.

The cDNA encoding heparanase is preferably cloned into a vector designed for expression in eukaryotic cells, rather than into a vector designed for expression in prokaryotic cells (e.g. *E. coli*). Eukaryotic cells are preferred for expression of genes obtained from higher eukaryotes because the signals for synthesis, processing, and secretion of these proteins are usually recognized, whereas this is often not true for prokaryotic hosts (Ausubel, *et al.*, ed., in Short Protocols in Molecular Biology, 2nd edition, John Wiley & Sons, publishers, pg.16-49, 1992.). Eukaryotic hosts may include, but are not limited to, the following: insect cells, African green monkey kidney cells (COS cells), Chinese hamster ovary cells (CHO cells), and Murine 3T3 fibroblasts.

Experiments demonstrating that a synthetic peptide of CTAP-III/NAP-2 or antisera raised against a synthetic peptide of CTAP-III/NAP-2 inhibit the heparanase activity of CTAP-III/NAP-2 suggest that the amino acids participating in enzymatic catalysis are contained in a C-terminal region of the enzyme.

<u>Peptide Synthesis:</u> A C-terminal peptide contained within the sequences known for CTAP-III (SEQ ID NO: 1), Platelet Basic Protein (SEQ ID NO: 3), β-thromboglobulin (SEQ ID NO: 5), and NAP-2 (SEQ ID NO: 7), and an N-terminal peptide contained within the sequences known for CTAP-III (SEQ ID NO: 1) and Platelet Basic Protein (SEQ ID NO: 3) were synthesized according to standard procedures. The N-terminal peptide has the following sequence (SEQ ID NO: 41: Asn Leu Ala Lys Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Cys, in which the final Cys residue was added to regions of known sequence (SEQ ID NOS: 1,3) for the purpose of

conjugation to a carrier protein. The C-terminal peptide has the following sequence (SEQ ID NO: 42): Cys Asn Gln Val Glu Val Ile Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp Pro Asp Ala Pro Arg Ile Lys Lys Ile Val Gln Lys Lys encoded by the cDNA sequence (SEQ ID NO: 43) of TGCAACCAAG TCGAAGTGAT AGCCACACTG AAGGATGGGA GGAAAATCTG CCTGGACCCA GATGCTCCCA GAATCAAGAA AATTGTACAG AAAAAA. These peptides (SEQ ID NOS: 41 and 42) were produced by stepwise solid phase peptide synthesis on an Applied Biosystems 430A Peptide Synthesizer. The 9-fluoroenylmethyloxycarbonyl (Fmoc) group was used as the N^{α} amino protecting group, and temporary side-chain protectin groups were as follows: Arg (Pmc), Asn (Trt), Asp (OtBu), Gln (Trt), Glu (OtBu), His (Trt), Lys (Boc), Ser (tBu), Thr (tBu). Each residue was single coupled using a HBTU/NMP protocol and capped with acetic anhydride before the next synthesis cycle. After removal of the N-terminal Fmoc group, temporary side-chain protecting groups were removed and the peptide cleaved from the resin by treatment with 95% TFA/5% scavengers (ethyl methyl sulfide/anisole/1,2-ethanedithiol, 1:3:1) for two hours at room temperature. The crude peptides were precipitated from the cleavage solution with cold diethyl ether. The precipitated peptide was collected on a sintered glass funnel, washed with diethyl ether, dissolved in dilute acetic acid, evaporated to dryness under reduced pressure, and the residue was redissolved and lyophillized from glacial acetic acid. The crude peptides were purified by preparative reverse phase chromatography on a Phenomenex C-18 column (22.5 x 250 mm) using a water/acetonitrile gradient, each phase containing 0.1% trifluoracetic acid (TFA). Clean fractions, as determined by analytical HPLC, were pooled, the acetonitrile was evaporated under reduced pressure, and the aqueous solution was lyophillized. The purified peptides were characterized by time of flight or FAB mass spectroscopy.

Further, SEQ ID NO: 42 can be produced by recombinant DNA methodology as stated in Procedure A (page 21).

Antisera Production: The synthetic peptides of CTAP-III/NAP-2 were conjugated to keyhole limpet hemocyanin utilizing a maleimide-activated carrier protein (Pierce Chemical Co. #77107). 300 µg of conjugated peptides were injected into chickens using Freunds complete adjuvant. The antisera were collected 5 weeks after initial immunization. Specific recognition by the antisera of commercial CTAP-III (2.5 µg, (Celsus Laboratories Inc., Cincinnati, Ohio; Cat #. 41705), isolated heparanase (1.5 µg), and 10 µl of the platelet supernatant used for purification was achieved by separating the proteins on a reducing 18% polyacrylamide gel (Novex), transferring to nitrocellulose, and incubating with the pre-immune or antisera (1:1500), followed by incubation with a peroxidase labeled goat anti-chicken IgG (1:500; Kierkegaard and Perry) in the presence of PBS containing 5% dry milk and 0.05% Tween-20. The pre-immune sera did not recognize 7 - 10 kD proteins in the commercial CTAP-III, isolated heparanase, or platelet

supernatants.

Inhibition of heparanase activity by the C-terminal synthetic peptide (SEQ ID NO: 42) or antisera: For experiments designed to determine whether the peptide antisera was able to inhibit heparanase activity, the pre-immune and antisera were exchanged into 0.15M NaCl, 0.01M sodium phoshate buffer, pH 7.4 (PBS) using a 100 kD cut-off membrane in order to remove low molecular weight chicken heparanase normally present in the serum. Aliquots of isolated heparanase (15 ng) were pre-incubated for 30 min with 2 µl of either pre-immune or anti-CTAP-III antisera before adding the ^{35}S-HSPG to determine heparanase activity. In the presence of the pre-immune sera, the isolated protein had 14.3 ± 0.1 units of heparanase activity, while in the presence of the C-terminal peptide antisera, only 0.8 ± 0.2 units of heparanase were detected ($p < 0.001$; results confirmed in a second experiment). The N-terminal peptide antiserum was not able to neutralize the heparanase activity. Similar results were obtained when the ability of the synthetic peptides to neutralize heparanase activity was examined. Heparanase assays conducted with 3 nM enzyme, 47 nM ^{35}S-HSPG substrate, and varying concentrations of peptides showed that heparanase activity was only 5% of control values in the presence of 250 µM C-terminal peptide. By contrast, heparanase activity in the presence of 250 µM of either the N-terminal peptide or an unrelated peptide (PLALWAR) was 67% of control values. The ability of both the C-terminal peptide (SEQ ID NO: 42) or antisera from a chicken immunized with the C-terminal synthetic peptide to neutralize heparanase activity demonstrates conclusively that CTAP-III and NAP-2 possess heparanase activity, and suggests that the C-terminal region is essential for catalysis. Modeling of this domain (SEQ ID NO: 42) can be used in the identification of potent peptide-mimetic compounds capable of inhibiting this enzyme activity.

Computer assisted modeling can be accomplished using programs for automated docking of molecules within 3D databases, as described in DesJarlais, R.L., Sheridan, R.P., Seibel, G.L., Dixon, J.S., Kuntz, I.D., Venkataraghavan, R., "Using shape complementarity as an initial screen in designing ligands for a receptor binding site of known three-dimensional structure"; J. Med. Chem. 31:722-729, 1988. Also, automated de novo construction of ligands that can bind the catalytic site as described in Moon, J.B., Howe, W.J., "Computer design of bioactive molecules: a method for receptor-based de novo ligand design"; Proteins: Struct., Funct., and Genetics, 11:314-328, 1981.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: Hoogewerf, Arlene J.
Ledbetter, Steven R.

(ii) TITLE OF INVENTION: USE OF HEPARANASE TO IDENTIFY AND ISOLATE ANTI-HEPARANASE COMPOUNDS

(iii) NUMBER OF SEQUENCES: 43

(iv) CORRESPONDENCE ADDRESS:
(A) ADDRESSEE: The Upjohn Company, Intellectual Property Law
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(C) CITY: Kalamazoo
(D) STATE: MI
(E) COUNTRY: USA
(F) ZIP: 49001

(v) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25

(vi) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(viii) ATTORNEY/AGENT INFORMATION:
(A) NAME: Jameson, William G.
(B) REGISTRATION NUMBER: 27,199
(C) REFERENCE/DOCKET NUMBER: 4731.1 CP

(ix) TELECOMMUNICATION INFORMATION:
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(B) TELEFAX: 616/385-6897
(C) TELEX: 224401

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 85 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

```
Asn Leu Ala Lys Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Tyr Ala
1               5                  10                  15

Glu Leu Arg Cys Met Cys Ile Lys Thr Thr Ser Gly Ile His Pro Lys
            20                  25                  30

Asn Ile Gln Ser Leu Glu Val Ile Gly Lys Gly Thr His Cys Asn Gln
        35                  40                  45

Val Glu Val Ile Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp
    50                  55                  60
```

Pro Asp Ala Pro Arg Ile Lys Lys Ile Val Gln Lys Lys Leu Ala Gly
65 70 75 80

Asp Glu Ser Ala Asp
85

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 255 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

AACTTGGCGA AAGGCAAAGA GGAAAGTCTA GACAGTGACT TGTATGCTGA ACTCCGCTGC 60

ATGTGTATAA AGACAACCTC TGGAATTCAT CCCAAAAACA TCCAAAGTTT GGAAGTGATC 120

GGGAAAGGAA CCCATTGCAA CCAAGTCGAA GTGATAGCCA CACTGAAGGA TGGGAGGAAA 180

ATCTGCCTGG ACCCAGATGC TCCCAGAATC AAGAAAATTG TACAGAAAAA ATTGGCAGGT 240

GATGAATCTG CTGAT 255

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 94 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Ser Ser Thr Lys Gly Gln Thr Lys Arg Asn Leu Ala Lys Gly Lys Glu
1 5 10 15

Glu Ser Leu Asp Ser Asp Leu Tyr Ala Glu Leu Arg Cys Met Cys Ile
20 25 30

Lys Thr Thr Ser Gly Ile His Pro Lys Asn Ile Gln Ser Leu Glu Val
35 40 45

Ile Gly Lys Gly Thr His Cys Asn Gln Val Glu Val Ile Ala Thr Leu
50 55 60

Lys Asp Gly Arg Lys Ile Cys Leu Asp Pro Asp Ala Pro Arg Ile Lys
65 70 75 80

Lys Ile Val Gln Lys Lys Leu Ala Gly Asp Glu Ser Ala Asp
85 90

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 282 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

TCCTCCACCA AAGGACAAAC TAAGAGAAAC TTGGCGAAAG GCAAAGAGGA AAGTCTAGAC 60

AGTGACTTGT ATGCTGAACT CCGCTGCATG TGTATAAAGA CAACCTCTGG AATTCATCCC 120

AAAAACATCC AAAGTTTGGA AGTGATCGGG AAAGGAACCC ATTGCAACCA AGTCGAAGTG 180

ATAGCCACAC TGAAGGATGG GAGGAAAATC TGCCTGGACC CAGATGCTCC CAGAATCAAG 240

AAAATTGTAC AGAAAAAATT GGCAGGTGAT GAATCTGCTG AT 282

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 81 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Tyr Ala Glu Leu Arg Cys
1 5 10 15

Met Cys Ile Lys Thr Thr Ser Gly Ile His Pro Lys Asn Ile Gln Ser
20 25 30

Leu Glu Val Ile Gly Lys Gly Thr His Cys Asn Gln Val Glu Val Ile
35 40 45

Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp Pro Asp Ala Pro
50 55 60

Arg Ile Lys Lys Ile Val Gln Lys Lys Leu Ala Gly Asp Glu Ser Ala
65 70 75 80

Asp

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 243 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

GGCAAAGAGG AAAGTCTAGA CAGTGACTTG TATGCTGAAC TCCGCTGCAT GTGTATAAAG 60

ACAACCTCTG GAATTCATCC CAAAAACATC CAAAGTTTGG AAGTGATCGG GAAAGGAACC 120

CATTGCAACC AAGTCGAAGT GATAGCCACA CTGAAGGATG GGAGGAAAAT CTGCCTGGAC 180

CCAGATGCTC CCAGAATCAA GAAAATTGTA CAGAAAAAAT TGGCAGGTGA TGAATCTGCT 240

GAT 243

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 69 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Glu Leu Arg Cys Met Cys Ile Lys Thr Thr Ser Gly Ile His Pro Lys
1 5 10 15

Asn Ile Gln Ser Leu Glu Val Ile Gly Lys Gly Thr His Cys Asn Gln
20 25 30

Val Glu Val Ile Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp
35 40 45

Pro Asp Ala Pro Arg Ile Lys Lys Ile Val Gln Lys Lys Leu Ala Gly
50 55 60

Asp Glu Ser Ala Asp
65

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 207 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

GAACTCCGCT GCATGTGTAT AAAGACAACC TCTGGAATTC ATCCCAAAAA CATCCAAAGT 60

TTGGAAGTGA TCGGGAAAGG AACCCATTGC AACCAAGTCG AAGTGATAGC CACACTGAAG 120

GATGGGAGGA AAATCTGCCT GGACCCAGAT GCTCCCAGAA TCAAGAAAAT TGTACAGAAA 180

AAATTGGCAG GTGATGAATC TGCTGAT 207

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 19 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Asn Leu Ala Lys Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Tyr Ala
1 5 10 15

Glu Leu Arg

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 16 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Ser Ser Thr Lys Gly Gln Thr Lys Arg Asn Leu Ala Lys Gly Lys Glu
1 5 10 15

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 17 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Asn Leu Ala Lys Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Tyr Ala
1 5 10 15

Glu

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 101 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Met Ser Ser Ala Ala Gly Phe Cys Ala Ser Arg Pro Gly Leu Leu Phe
1 5 10 15

Leu Gly Leu Leu Leu Leu Pro Leu Val Val Ala Phe Ala Ser Ala Glu
20 25 30

Ala Glu Glu Asp Gly Asp Leu Gln Cys Leu Cys Val Lys Thr Thr Ser
35 40 45

Gln Val Arg Pro Arg His Ile Thr Ser Leu Glu Val Ile Lys Ala Gly
50 55 60

Pro His Cys Pro Thr Ala Gln Leu Ile Ala Thr Leu Lys Asn Gly Arg
65 70 75 80

Lys Ile Cys Leu Asp Leu Gln Ala Pro Leu Tyr Lys Lys Ile Ile Lys
85 90 95

Lys Leu Leu Glu Ser
100

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 439 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

CCGCAGCATG AGCTCCGCAG CCGGGTTCTG CGCCTCACGC CCCGGGCTGC TGTTCCTGGG 60

GTTGCTGCTC CTGCCACTTG TGGTCGCCTT CGCCAGCGCT GAAGCTGAAG AAGATGGGGA 120

CCTGCAGTGC CTGTGTGTGA AGACCACCTC CCAGGTCCGT CCCAGGCACA TCACCAGCCT 180

GGAGGTGATC AAGGCCGGAC CCCACTGCCC CACTGCCCAA CTGATAGCCA CGCTGAAGAA 240

TGGAAGGAAA ATTTGCTTGG ACCTGCAAGC CCCGCTGTAC AAGAAAATAA TTAAGAAACT 300

TTTGGAGAGT TAGCTACTAG CTGCCTACGT GTGTGCATTT GCTATATAGC ATACTTCTTT 360

TTTCCAGTTT CAATCTAACT GTGAAAGAAA CTTCTGATAT TTGTGTTATC CTTATGATTT 420

TAAATAAACA AAATAAATC 439

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 101 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Met Asn Gln Thr Ala Ile Leu Ile Cys Cys Leu Ile Phe Leu Thr Leu
1 5 10 15

Ser Gly Ile Gln Gly Val Pro Leu Ser Arg Thr Val Arg Cys Thr Cys
20 25 30

Ile Ser Ile Ser Asn Gln Pro Val Asn Pro Val Asn Pro Arg Ser Leu
35 40 45

Glu Lys Leu Glu Ile Ile Pro Ala Ser Gln Phe Cys Pro Arg Val Glu
50 55 60

Ile Ile Ala Thr Met Lys Lys Lys Gly Glu Lys Arg Cys Leu Asn Pro
65 70 75 80

Glu Ser Lys Ala Ile Lys Asn Leu Leu Lys Ala Val Ser Lys Glu Met
85 90 95

Ser Lys Arg Ser Pro
100

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 650 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

GAGACATTCC TCAATTGCTT AGACATATTC TGAGCCTACA GCAGAGGAAC CTCCAGTCTC 60

AGCACCATGA ATCAAACTGC GATTCTGATT TGCTGCCTTA TCTTTCTGAC TCTAAGTGGC 120

ATTCAAGGAG TACCTCTCTC TAGAACCGTA CGCTGTACCT GCATCAGCAT TAGTAATCAA 180

CCTGTTAATC CAAGGTCTTT AGAAAAACTT GAAATTATTC CTGCAAGCCA ATTTTGTCCA 240

CGTGTTGAGA TCATTGCTAC AATGAAAAAG AAGGGTGAGA AGAGATGTCT GAATCCAGAA 300

TCGAAGGCCA TCAAGAATTT ACTGAAAGCA GTTAGCAAGG AAATGTCTAA AAGATCTCCT 360

TAAAACCAGA GGGGAGCAAA ATCGATGCAG TGCTTCCAAG GATGGACCAC ACAGAGGCTG 420

CCTCTCCCAT CACTTCCCTA CATGGAGTAT ATGTCAAGCC ATAATTGTTC TTAGTTTGCA 480

GTTACACTAA AAGGTGACCA ATGATGGTCA CCAAATCAGC TGCTACTACT CCTGTAGGAA 540

GGTTAATGTT CATCATCCTA AGCTATTCAG TAATAACTCT ACCCTGGCAC TATAATGTAA 600

GCTCTACTGA GGTGCTATGT TCTTAGTGGA TGTTCTGACC CTGCTTCAAA 650

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 107 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Met Ala Arg Ala Ala Leu Ser Ala Ala Pro Ser Asn Pro Arg Leu Leu
1 5 10 15

Arg Val Ala Leu Leu Leu Leu Leu Leu Val Ala Ala Gly Arg Arg Ala
20 25 30

Ala Gly Ala Ser Val Ala Thr Glu Leu Arg Cys Gln Cys Leu Gln Thr
35 40 45

Leu Gln Gly Ile His Pro Lys Asn Ile Gln Ser Val Asn Val Lys Ser
50 55 60

Pro Gly Pro His Cys Ala Gln Thr Glu Val Ile Ala Thr Leu Lys Asn
65 70 75 80

Gly Arg Lys Ala Cys Leu Asn Pro Ala Ser Pro Ile Val Lys Lys Ile
85 90 95

Ile Glu Lys Met Leu Asn Ser Asp Lys Ser Asn
100 105

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1050 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

CTCGCCAGCT CTTCCGCTCC TCTCACAGCC GCCAGACCCG CCTGCTGAGC CCCATGGCCC 60

GCGCTGCTCT CTCCGCCGCC CCCAGCAATC CCCGGCTCCT GCGAGTGGCA CTGCTGCTCC 120

TGCTCCTGGT AGCCGCTGGC CGGCGCGCAG CAGGAGCGTC CGTGGCCACT GAACTGCGCT 180

GCCAGTGCTT GCAGACCCTG CAGGGAATTC ACCCCAAGAA CATCCAAAGT GTGAACGTGA 240

AGTCCCCCGG ACCCCACTGC GCCCAAACCG AAGTCATAGC CACACTCAAG AATGGGCGGA 300

AAGCTTGCCT CAATCCTGCA TCCCCCATAG TTAAGAAAAT CATCGAAAAG ATGCTGAACA 360

GTGACAAATC CAACTGACCA GAAGGGAGGA GGAAGCTCAC TGGTGGCTGT TCCTGAAGGA 420

GGCCCTGCCC TTATAGGAAC AGAAGAGGAA AGAGAGACAC AGCTGCAGAG GCCACCTGGA 480

TTGTGCCTAA TGTGTTTGAG CATCGCTTAG GAGAAGTCTT CTATTTATTT ATTTATTCAT 540

TAGTTTTGAA GATTCTATGT TAATATTTTA GGTGTAAAAT AATTAAGGGT ATGATTAACT 600

CTACCTGCAC ACTGTCCTAT TATATTCATT CTTTTTGAAA TGTCAACCCC AAGTTAGTTC 660

AATCTGGATT CATATTTAAT TTGAAGGTAG AATGTTTTCA AATGTTCTCC AGTCATTATG 720

TTAATATTTC TGAGGAGCCT GCAACATGCC AGCCACTGTG ATAGAGGCTG GCGGATCCAA 780

GCAAATGGCC AATGAGATCA TTGTGAAGGC AGGGGAATGT ATGTGCACAT CTGTTTTGTA 840

ACTGTTTAGA TGAATGTCAG TTGTTATTTA TTGAAATGAT TTCACAGTGT GTGGTCAACA 900

TTTCTCATGT TGAAACTTTA AGAACTAAAA TGTTCTAAAT ATCCCTTGGA CATTTTATGT 960

CTTTCTTGTA AGGCATACTG CCTTGTTTAA TGGTAGTTTT ACAGTGTTTC TGGCTTAGAA 1020

CAAAGGGGCT TAATTATTGA TGTTTTCGGA 1050

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 102 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Met Ala Arg Ala Thr Leu Ser Ala Ala Pro Ser Asn Pro Arg Leu Leu
1 5 10 15

Arg Val Ala Leu Leu Leu Leu Leu Leu Val Ala Ala Ser Arg Arg Ala
20 25 30

Ala Gly Ala Pro Lys Ala Thr Glu Lys Arg Cys Gln Cys Lys Gln Thr
35 40 45

Leu Gln Gly Ile His Leu Lys Asn Ile Gln Ser Val Lys Val Lys Ser
50 55 60

Pro Gly Pro His Cys Ala Gln Thr Glu Val Ile Ala Thr Leu Lys Asn
65 70 75 80

Gly Gln Lys Ala Cys Leu Asn Pro Ala Ser Pro Met Val Lys Lys Ile
85 90 95

Ile Glu Lys Met Leu Lys
100

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1081 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

CTCTCCTCCT CGCACAGCCG CTCGAACCGC CTGCTGAGCC CCATGGCCCG CGCCACGCTC 60

TCCGCCGCCC CCAGCAATCC CCGGCTCCTG CGGGTGGCGC TGCTGCTCCT GCTCCTGGTG 120

GCCGCCAGCC GGCGCGCAGC AGGAGCGCCC CTGGCCACTG AACTGCGCTG CCAGTGCTTG 180

CAGACCCTGC AGGGAATTCA CCTCAAGAAC ATCCAAAGTG TGAAGGTGAA GTCCCCCGGA 240

CCCCACTGCG CCCAAACCGA AGTCATAGCC ACACTCAAGA ATGGGCAGAA AGCTTGTCTC 300

AACCCCGCAT CGCCCATGGT TAAGAAAATC ATCGAAAAGA TGCTGAAAAA TGGCAAATCC 360

AACTGACCAG AAGGAAGGAG GAAGCTTATT GGTGGCTGTT CCTGAAGGAG GCCCTGCCCT 420

TACAGGAACA GAAGAGGAAA GAGAGACACA GCTGCAGAGG CCACCTGGAT TGCGCCTAAT 480

GTGTTTGAGC ATCACTTAGG AGAAGTCTTC TATTTATTTA TTTATTTATT TATTTGTTTG 540

TTTTAGAAGA TTCTATGTTA ATATTTTATG TGTAAAATAA GGTTATGATT GAATCTACTT 600

GCACACTCTC CCATTATATT TATTGTTTAT TTTAGGTCAA ACCCAAGTTA GTTCAATCCT 660

GATTCATATT TAATTTGAAG ATAGAAGGTT TGCAGATATT CTCTAGTCAT TTGTTAATAT 720

TTCTTCGTGA TGACATATCA CATGTCAGCC ACTGTGATAG AGGCTGAGGA ATCCAAGAAA 780

ATGGCCAGTG AGATCAATGT GACGGCAGGG AAATGTATGT GTGTCTATTT TGTAACTGTA 840

AAGATGAATG TCAGTTGTTA TTTATTGAAA TGATTTCACA GTGTGTGGTC AACATTTCTC 900

ATGTTGAAGC TTTAAGAACT AAAATGTTCT AAATATCCCT TGGACATTTT ATGTCTTTCT 960

TGTAAGGCAT ACTGCCTTGT TTAATGTTAA TTATGCAGTG TTTCCCTCTG TGTTAGAGCA 1020

GAGAGGTTTC GATATTTATT GATGTTTTCA CAAAGAACAG GAAAATAAAA TATTTAAAAA 1080

T 1081

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 107 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Met Ala His Ala Thr Leu Ser Ala Ala Pro Ser Asn Pro Arg Leu Leu
1 5 10 15

Arg Val Ala Leu Leu Leu Leu Leu Leu Val Ala Ala Ser Arg Arg Ala
20 25 30

Ala Gly Ala Ser Val Val Thr Glu Leu Arg Cys Gln Cys Leu Gln Thr
35 40 45

Leu Gln Gly Ile His Leu Lys Asn Ile Gln Ser Val Asn Val Arg Ser
50 55 60

Pro Gly Pro His Cys Ala Gln Thr Glu Val Ile Ala Thr Leu Lys Asn
65 70 75 80

Gly Lys Lys Ala Cys Leu Asn Pro Ala Ser Pro Met Val Gln Lys Ile
85 90 95

Ile Glu Lys Ile Leu Asn Lys Gly Ser Thr Asn
100 105

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 988 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

CTCGCACAGC TTCCCGACGC GTCTGCTGAG CCCCATGGCC CACGCCACGC TCTCCGCCGC 60

CCCCAGCAAT CCCCGGCTCC TGCGGGTGGC GCTGCTGCTC CTGCTCCTGG TGGCCGCCAG 120

CCGGCGCGCA GCAGGAGCGT CCGTGGTCAC TGAACTGCGC TGCCAGTGCT TGCAGACACT 180

GCAGGGAATT CACCTCAAGA ACATCCAAAG TGTGAATGTA AGGTCCCCCG GACCCCACTG 240

CGCCCAAACC GAAGTCATAG CCACACTCAA GAATGGGAAG AAAGCTTGTC TCAACCCCGC 300

ATCCCCCATG GTTCAGAAAA TCATCGAAAA GATACTGAAC AAGGGGAGCA CCAACTGACA 360

GGAGAGAAGT AAGAAGCTTA TCAGCGTATC ATTGACACTT CCTGCAGGGT GGTCCCTGCC 420

CTTACCAGAG CTGAAAATGA AAAAGAGAAC AGCAGCTTTC TAGGGACAGC TGGAAAGGAC 480

TTAATGTGTT TGACTATTTC TTACGAGGGT TCTACTTATT TATGTATTTA TTTTTGAAAG 540

CTTGTATTTT AATATTTTAC ATGCTGTTAT TTAAAGATGT GAGTGTGTTT CATCAAACAT 600

AGCTCAGTCC TGATTATTTA ATTGGAATAT GATGGGTTTT AAATGTGTCA TTAAACTAAT 660

ATTTAGTGGG AGACCATAAT GTGTCAGCCA CCTTGATAAA TGACAGGGTG GGGAACTGGA 720

GGGTGGGGGG ATTGAAATGC AAGCAATTAG TGGATCACTG TTAGGGTAAG GGAATGTATG 780

TACACATCTA TTTTTTATAC TTTTTTTTTA AAAAAAGAAT GTCAGTTGTT ATTTATTCAA 840

ATTATCTCAC ATTATGTGTT CAACATTTTT ATGCTGAAGT TTCCCTTAGA CATTTTATGT 900

CTTGCTTGTA GGGCATAATG CCTTGTTTAA TGTCCATTCT GCAGCGTTTC TCTTTCCCTT 960

GGAAAAGAGA ATTTATCATT ACTGTTAC 988

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 97 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Met Thr Ser Lys Leu Ala Val Ala Leu Leu Ala Ala Phe Leu Ile Ser
1 5 10 15

Ala Ala Leu Cys Glu Gly Ala Val Leu Pro Arg Ser Ala Lys Glu Leu
20 25 30

Arg Cys Gln Cys Ile Lys Thr Tyr Ser Lys Pro Phe His Pro Lys Phe
35 40 45

Ile Lys Glu Leu Arg Val Ile Glu Ser Gly Pro His Cys Ala Asn Thr
50 55 60

Glu Ile Ile Val Lys Leu Ser Asp Gly Arg Glu Leu Cys Leu Asp Pro
65 70 75 80

Lys Glu Asn Trp Val Gln Arg Val Val Glu Lys Phe Leu Lys Arg Ala
85 90 95

Glu

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 291 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

ATGACTTCCA AGCTGGCCGT GGCTCTCTTG GCAGCCTTCC TGATTTCTGC AGCTCTGTGT 60

GAAGGTGCAG TTTTGCCAAG GAGTGCTAAA GAACTTAGAT GTCAGTGCAT AAAGACATAC 120

TCCAAACCTT TCCACCCCAA ATTTATCAAA GAACTGAGAG TGATTGAGAG TGGACCACAC 180

TGCGCCAACA CAGAAATTAT TGTAAAGCTT TCTGATGGAA GAGAGCTCTG TCTGGACCCC 240

AAGGAAAACT GGGTGCAGAG GGTTGTGGAG AAGTTTTTGA AGAGGGCTGA G 291

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 78 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Ala Gly Pro Ala Ala Ala Val Leu Arg Glu Lys Arg Cys Val Cys Leu
1 5 10 15

Gln Thr Thr Gln Gly Val His Pro Lys Met Ile Ser Asn Leu Gln Val
20 25 30

Phe Ala Ile Gly Pro Gln Cys Ser Lys Val Glu Val Val Ala Ser Leu
35 40 45

Lys Asn Gly Lys Glu Ile Cys Leu Asp Pro Glu Ala Pro Phe Leu Lys
50 55 60

Lys Val Ile Gln Lys Ile Leu Asp Gly Gly Asn Lys Glu Asn
65 70 75

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 216 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

GTGTTGCGGG AACTGCGGTG CGTGTGTTTA CAGACCACGC AGGGAGTTCA TCCCAAAATG 60

ATCAGTAATC TGCAAGTGTT CGCCATAGGC CCACAGTGCT CCAAGGTGGA AGTGGTAGCC 120

TCCCTGAAGA ACGGGAAGGA AATTTGTCTT GATCCAGAAG CCCCTTTTCT AAAGAAAGTC 180

ATCCAGAAAA TCCTCGACGG CGGCAACAAA GAAAAC 216

(2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 93 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Met Gln Val Ser Thr Ala Ala Leu Ala Val Leu Leu Cys Thr Met Ala
1 5 10 15

Leu Cys Asn Gln Val Leu Ser Ala Pro Leu Ala Ala Asp Thr Pro Thr
20 25 30

Ala Cys Cys Phe Ser Tyr Thr Ser Arg Gln Ile Pro Gln Asn Phe Ile
35 40 45

Ala Asp Tyr Phe Glu Thr Ser Ser Gln Cys Ser Lys Pro Ser Val Ile
50 55 60

Phe Leu Thr Lys Arg Gly Arg Gln Val Cys Ala Asp Pro Ser Glu Glu
65 70 75 80

Trp Val Gln Lys Tyr Val Ser Asp Leu Glu Leu Ser Ala
85 90

(2) INFORMATION FOR SEQ ID NO:27:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 4788 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

GAATTCAAGG CCTGTCCTGG TTTGGTCCCA ATTTACCTTT ATCATCCATA TTCACCCCCA 60

CTGCTCTGCA GCTCCACTGA AGCACCCCCT CTTTCCTCTG AGCCACAATG TCACACCCAG 120

GACTCTGCCT CAGCTGGGCC TCCACTGCCC ACCCATCTAT AGATGCCTAA ATCCCGGGCA 180

GTTATCCAGA CACAACTAAA GTTCCATCCC TTCCATGAAG CCTTCCCCAA CCCTCTGGTG 240

GAAGGTCACT TCTTCCTCAT GGGGTTCTGA GCTTTCATTT CTTTTTCTAC TAAGAGTTTT 300

ACAATTACCT GTTCATACAC TCTACCTGCC CCCATGAGAC CAGGGGCATC TCAGAAACAA 360

AGATCATTAA AACCAACTAA ATCTATTTCT CATTATAAAA TGAGATATGC TGATTGATTG 420

CAAAATAATA AAATAACAAA GTATGGAAAA GAAAAAAAAA AGCATATAAT CTGGCTGAGA 480

AGGTAGAGAC CCTTCCACAC CACTGAAATT ATGTGTTGAA AAGAATAAGG AAAAAACTGC 540

TTCAGTTTGG CATTATTTAT GTAAGTATAG TATAGGATCC TTAAAATGGT TCAAAGAAAT 600

GGGAAATCAA GACTTCATTT TGGCAAAGCC ATTGAACAGA AACTGTAGCA TATTTATCAG 660

TAATTTCTTT CAGATTAAAC AACTGACAAC AACCCACTTT TCAACCAGTG ATGTTGGAAA 720

TGTTTTAAAA CAAAATTAGT TCATAAATTT GTGGGTTGAC CAAGAAGGTA ATAAAGTCTC 780

ACTAAATAAA ATGAGGAAAA TTCAGAAAAA GAAAAAAATA AGAAAATAAA TCACCCATGG 840

ATCTAAGCAC TATTCATTCT TTAAGGCATG TATTTCCAAG CCTTTTAATT TTTTCATGCC 900

TAGAGTTGGC ATGGCATATA TATATCTTTA TACAATTCTT CAAATTTTAT AGAATTTGTA 960

TAATGTTTTA TCTTGCTTTT TTTTTAACCA CTGATGTTAT AAGCATATTT ATGCCACTTC 1020

ATTCACGTTA GAGACTTAAT AATAAAGGAT CTTGTGGATA ATTTATCATT CCCTGATAGA 1080

GAAAAATTTA GCTTTGCTTA TTTTAGAGTT ATAAATGATG CTGGGTCAGG TATCTTTATG 1140

TTTGAAGATG GCTCCATATT TGGGTTGTTT CCACAGAACT CTTTCCAGAA ATGCTTTTTC 1200

TAGGTTAATG GCTACACATA TTTCTAGGCA CCTGACATAC TGACACCCAC CTCTAAAGTA 1260

TTTTTATGAT CCACAACTAG CGTTTAACAC AGCGCCCCAG TCACTCCGAG ACTAATAAAT 1320

AGACAAATGA CTGAAACGTG ACCTCATGCT TTCTATTCCT CCAGCTTTCA TTGAGTTCCT 1380

TTCCTCTGGG AGGACTGGGG GTTGTCTAGC CCTCCACAGC ATCAGCCCAT TGACCCTATC 1440

CTTGTGGTTA TAGCAGCTGA GGAAGCAGAA TTACAGCTCT GTGGGAAGGA ATGGGGCTGG 1500

AGAGTTCATG CATAGACCAA TTCTTTTTTT TTTTTTTTTT TGAGATGGAG TTTCACTTTT 1560

GTTGCCCAGG CTGGAGTGCA ATGGCATGAT CTCAGCTCAC CACAGCCCCC ACCTCCTGGG 1620

TTCAAGCGAT TCTCCTGCCC TCAGCCTCCC GAGTAGCTGG GATTACAGGC ATGTGCCACC 1680

ACGCCTGACT ACTTTTGTAT TTTTAGTAGA GATGGAGTTT CTCTTTCTTG GTCAGGTTGG 1740

TCTCAAACTC CTGACCTCAG GTGATCTGCA GCCTCGGCCT CCAAAGTGTT GGGATTACAG 1800

GTGTGAGCGA CCATGCCTGG CTGCATAGAC CAGTTCTTAT GAGAAGGGAT CAACTAAGAA 1860

TAGCCTTGGG TTGACACACA CCCCTCTTCA CACTCACAGG AGAAACCCCA TGAAGCTAGA 1920

ACCAGTCATG AGTTGAGAGC TGAGAGTTAG AGAGTAGCTC AGAGATGCTA TTCTTGGATA 1980

TCCTGAGCCC CTGTGGTCAC CAGGGACCCT GAGTTGTGCA ACACTCAGCA TGACAGCATC 2040

ACTACACTTA AAAATTTCCC TCCTCACCCC CAGATTCCAT TTCCCCATCC GCCAGGGCTG 2100

CCTATAAAGA GGAGAGATGG CTTCAGACAT CAGAAGGACG CAGGCAGCAA AGAGTAGTCA 2160

GTCCCTTCTT GGCTCTGCTG ACACTCGAGC CCACATTCCA TCACCTGCTC CCAATCATGC 2220

AGGTCTCCAC TGCTGCCCTT GCCGTCCTCC TCTGCACCAT GGCTCTCTGC AACCAGGTCC 2280

TCTCTGCACC ACGTGAGTCC ATGTTGTTGT TGTGGGTATC ACCACTCTCT GGCCATGGTT 2340

AGACCACATC AGTCTTTTTT TGTGGCGTGA GAGGCCCCGA AGAGAAAAGA AGGAAGTTCT 2400

TAAAGCGCTG CCAAACACCT TGGTCTTTTT CTTCACAACT TTTATTTTTA TCTCTAGAAG 2460

GGGTCTTAGC CCTCCTAGTC TCCAGGTATG AGAATCTAGG CAGGGGCAGG GGAGTTACAG 2520

TCCCTTGTAC AGATAGAAAA ACAGGGTTCA AAACGAATCA GTTTGCAAGA GGCAGAATCC 2580

AGGGCTGCTT ACTTCCCAGT GGGGTCTGTT CTTCACTCTC CAGCTCACCC TAGTCTCCCA 2640

GGAGCCCTGT CCCTTGGATG TCTTATGAGA GATGTCCAGG GCTTCTCTTG GGCTGGGGTA 2700

TGACTTCTTG AACCGACAAA ATTCCATGAA GAGAGCTAAG AGAACAGTCC ATTCAGGTAT 2760

CTGGATCACA TAGAGAAACA GAGAACCCAC TATGAAGAGT CAAGGGGAAA GAGGAATATA 2820

GACAGAAACA AAGAGACATT TCTCTGCAAA ACCCCCCAAA TGCCTTGCAG TCACTTGGTC 2880

TGAGCAAGCC TGCCCTCCTC AACCACTCAG GGATCAGAAG CTGCCTGGCC TTTTCTTCTG 2940

AGCTGTGACT TGGGCTTATT CTCTCCTTTC TCCGCAGTTG CTGCTGACAC GCCGACCGCC 3000

TGCTGCTTCA GCTACACCTC CCGACAGATT CCACAGAATT TCATAGCTGA CTACTTTGAG 3060

ACGAGCAGCC AGTGCTCCAA GCCCAGTGTC ATGTAAGTGC CAGTCTTCCT GCTCACCTCT 3120

AGGGAGGTAG GGAGTGTCAG GGTGGGGGCA GAAACAGGCC AGAAGGCCAT CCTGGAAAGG 3180

CCCAGCCTTC AGGAGCCTAT CGGGGATACA GGACGCAGGG CACTGAGGTG TGACCTGACT 3240

TGGGGCTGGA GTGAGGTGGG TGTTACAGAG TCAGGAAGGG CTGCCCCAGG CCAGAGGAAA 3300

GGGACAGGAA GAAGGAGGCA GCAGGACACT CTGAGGGCCC CCTTGCCTGG AGTCACTGAG 3360

AGAAGCTCTC TAGACGGAGA TAGGCAGGGG GCCCCTGAGA GAGGAGCAGG CCTTGAGCTG 3420

CCCAGGACAG AGAGCAGGAT GTCAGGGCCA TGGTGGGCCC AGGATTCCCC GGCTGGATTC 3480

CCCAGTGCTT AACTCTTCCT CCCTTCTCCA CAGCTTCCTA ACCAAGAGAG GCCGGCAGGT 3540

CTGTGCTGAC CCCAGTGAGG AGTGGGTCCA GAAATACGTC AGTGACCTGG AGCTGAGTGC 3600

CTGAGGGGTC CAGAAGCTTC GAGGCCCAGC GACCTCAGTG GGCCCAGTGG GGAGGAGCAG 3660

GAGCCTGAGC CTTGGGAACA TGCGTGTGAC CTCCACAGCT ACCTCTTCTA TGGACTGGTT 3720

ATTGCCAAAC AGCCACACTG TGGGACTCTT CTTAACTTAA ATTTTAATTT ATTTATACTA 3780

TTTAGTTTTT ATAATTTATT TTTGATTTCA CAGTGTGTTT GTGATTGTTT GCTCTGAGAG 3840

TTCCCCCTGT CCCCTCCACC TTCCCTCACA GTGTGTCTGG TGACAACCGA GTGGCTGTCA 3900

TCGGCCTGTG TAGGCAGTCA TGGCACCAAA GCCACCAGAC TGACAAATGT GTATCAGATG 3960

CTTTTGTTCA GGGCTGTGAT CGGCCTGGGG AAATAATAAA GATGTTCTTT TAAACGGTAA 4020

ACCAGTATTG AGTTTGGTTT TGTTTTTCTG GCAAATCAAA ATCACTGGTT AAGAGGAATC 4080

ATAGGCAAAG ATTAGGAAGA GGTGAAATGG AGGGAAATTG GGAGAGATGG GGAGCGCTGC 4140

GACAGAGTTA TCCACTTCAC AAAATTCTGG AACATTGAAA CTACGAATAT GTTATAACTC 4200

AAATCGTAAT ATGCACGCTC TAGGAGAATT AACTACTTGA ATGGCCACCA TTAAGCAGAG 4260

TATTCTGTAG GGCATATTCA TGATGAATCA AGCTCTTAAT AGCAATTATT TACATTGTTG 4320

AGGCTTACTC CTCCTACTGA GTGCTTTTTA TACATTGTTC ATTTAATCTT ACCAATGCAA 4380

TAGTACAGCT TAGGTACTAT TAATACCTCC ACTTGACAGA AAAGTAACCC AGGGCTCAGA 4440

AAGGTTAGAC AACTTGGCTG AGGTTACACA GCACGTAAAC GGTCAATTGT GTTCCAAAAC 4500

TGGACTTTTA TTGAACTACA GACTATGCTG TTAACCATTG ACCAAGTTAT TTCCCAAAGT 4560

ATGACCCGCC TATACTCAAA TCTTACCCCA TTCTTTAACA GATGATACTT TATCCATTGC 4620

AACCACTTCC TGTCAGGATT CTGAGTTGAC ATAGAGTGTT TCAGCAGTGA TTATTTAAGC 4680

CAATTACATC AGGATCTTTA GGTGTAGACC TGGGAACTGA TATTTTTATC AAGCTCATGA 4740

GGTGTTCCAT AGCATGTTAA TGACTGAGAG CCACTGTCAA TAGAATTC 4788

(2) INFORMATION FOR SEQ ID NO:28:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 92 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Met Lys Leu Cys Val Thr Val Leu Ser Leu Leu Met Leu Val Ala Ala
1 5 10 15

Phe Cys Ser Pro Ala Leu Ser Ala Pro Met Gly Ser Asp Pro Pro Thr
20 25 30

Ala Cys Cys Phe Ser Tyr Thr Ala Arg Lys Leu Pro Arg Asn Phe Val
35 40 45

Val Asp Tyr Tyr Glu Thr Ser Ser Leu Cys Ser Gln Pro Ala Val Val
50 55 60

Phe Gln Thr Lys Arg Ser Lys Gln Val Cys Ala Asp Pro Ser Glu Ser
65 70 75 80

Trp Val Gln Glu Tyr Val Tyr Asp Leu Glu Leu Asn
85 90

(2) INFORMATION FOR SEQ ID NO:29:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 696 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

TTCCCCCCCC CCCCCCCCCC CCCCGCCCGA GCACAGGACA CAGCTGGGTT CTGAAGCTTC 60

TGAGTTCTGC AGCCTCACCT CTGAGAAAAC CTCTTTTCCA CCAATACCAT GAAGCTCTGC 120

GTGACTGTCC TGTCTCTCCT CATGCTAGTA GCTGCCTTCT GCTCTCCAGC GCTCTCAGCA 180

CCAATGGGCT CAGACCCTCC CACCGCCTGC TGCTTTTCTT ACACCGCGAG GAAGCTTCCT 240

CGCAACTTTG TGGTAGATTA CTATGAGACC AGCAGCCTCT GCTCCCAGCC AGCTGTGGTA 300

TTCCAAACCA AAAGAAGCAA GCAAGTCTGT GCTGATCCCA GTGAATCCTG GGTCCAGGAG 360

GGGCTCTGGA AACCACATGG CTTCACCTGT CCCCGAAACT ACCAGCCCTA CACCATTCCT 420

TCTGCCCTGC TTTTGCTAGG TCACAGAGGA TCTGCTTGGT CTTGATAAGC TATGTTGTTG 480

CACTTTAAAC ATTTAAATTA TACAATCATC AACCCCCAAC 520

(2) INFORMATION FOR SEQ ID NO:32:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 99 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Met Lys Val Ser Ala Ala Leu Leu Cys Leu Leu Leu Ile Ala Ala Thr
1 5 10 15

Phe Ile Pro Gln Gly Lys Ala Gln Pro Asp Ala Ile Asn Ala Pro Val
20 25 30

Thr Cys Cys Tyr Asn Phe Thr Asn Arg Lys Ile Ser Val Gln Arg Leu
35 40 45

Ala Ser Tyr Arg Arg Ile Thr Ser Ser Lys Cys Pro Lys Glu Ala Val
50 55 60

Ile Phe Lys Thr Ile Val Ala Lys Glu Ile Cys Ala Asp Pro Lys Gln
65 70 75 80

Lys Trp Val Gln Asp Ser Met Asp His Leu Asp Lys Gln Thr Gln Thr
85 90 95

Pro Lys Thr

(2) INFORMATION FOR SEQ ID NO:33:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 725 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

CTAACCCAGA AACATCCAAT TCTCAAACTG AAGCTCGCAC TCTCGCCTCC AGCATGAAAG 60

TCTCTGCCGC CCTTCTGTGC CTGCTGCTCA TAGCAGCCAC CTTCATTCCC CAAGGGCTCG 120

CTCAGCCAGA TGCAATCAAT GCCCCAGTCA CCTGCTGTTA TAACTTCACC AATAGGAAGA 180

TCTCAGTGCA GAGGCTCGCG AGCTATAGAA GAATCACCAG CAGCAAGTGT CCCAAAGAAG 240

CTGTGATCTT CAAGACCATT GTGGCCAAGG AGATCTGTGC TGACCCCAAG CAGAAGTGGG 300

TTCAGGATTC CATGGACCAC CTGGACAAGC AAACCCAAAC TCCGAAGACT TGAACACTCA 360

CTCCACAACC CAAGAATCTG CAGCTAACTT ATTTTCCCCT AGCTTTCCCC AGACACCCTG 420

TTTTATTTTA TTATAATGAA TTTTGTTTGT TGATGTGAAA CATTATGCCT TAAGTAATGT 480

TAATTCTTAT TTAAGTTATT GATGTTTTAA GTTTATCTTT CATGGTACTA GTGTTTTTTA 540

GATACAGAGA CTTGGGGAAA TTGCTTTTCC TCTTGAACCA CAGTTCTACC CCTGGGATGT 600

TTTGAGGGTC TTTGCAAGAA TCATTAATAC AAAGAATTTT TTTTAACATT CCAATGCATT 660

GCTAAAATAT TATTGTGGAA ATGAATATTT TGTAACTATT ACACCAAATA AATATATTTT 720

TGTAC 725

(2) INFORMATION FOR SEQ ID NO:34:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 99 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Met Lys Ala Ser Ala Ala Leu Leu Cys Leu Leu Leu Thr Ala Ala Ala
1 5 10 15

Phe Ser Pro Gln Gly Leu Ala Gln Pro Val Gly Ile Asn Thr Ser Thr
20 25 30

Thr Cys Cys Tyr Arg Phe Ile Asn Lys Lys Ile Pro Lys Gln Arg Leu
35 40 45

Glu Ser Tyr Arg Arg Thr Thr Ser Ser His Cys Pro Arg Glu Ala Val
50 55 60

Ile Phe Lys Thr Lys Leu Asp Lys Glu Ile Cys Ala Asp Pro Thr Gln
65 70 75 80

Lys Trp Val Gln Asp Phe Met Lys His Leu Asp Lys Lys Thr Gln Thr
85 90 95

Pro Lys Leu

(2) INFORMATION FOR SEQ ID NO:35:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 810 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

AGCAGAGGGG CTGAGACCAA ACCAGAAACC TCCAATTCTC ATGTGGAAGC CCATGCCCTC 60

ACCCTCCAAC ATGAAAGCCT CTGCAGCACT TCTGTGTCTG CTGCTCACAG CAGCTGCTTT 120

CAGCCCCCAG GGGCTTGCTC AGCCAGTTGG GATTAATACT TCAACTACCT GCTGCTACAG 180

ATTTATCAAT AAGAAAATCC CTAAGCAGAG GCTGGAGAGC TACAGAAGGA CCACCAGTAG 240

CCACTGTCCC CGGGAAGCTG TAATCTTCAA GACCAAACTG GACAAGGAGA TCTGTGCTGA 300

CCCCACACAG AAGTGGGTCC AGGACTTTAT GAAGCACCTG GACAAGAAAA CCCAAACTCC 360

AAAGCTTTGA ACATTCATGA CTGAACTAAA AACAAGCCAT GACTTGAGAA ACAAATAATT 420

TGTATACCCT GTCCTTTCTC AGAGTGGTTC TGAGATTATT TTAATCTAAT TCTAAGGAAT 480

ATGAGCTTTA TGTAATAATG TGAATCATGG TTTTTCTTAG TAGATTTTAA AAGTTATTAA 540

TATTTTAATT TAATCTTCCA TGGATTTTGG TGGGTTTTGA ACATAAAGCC TTGGATGTAT 600

ATGTCATCTC AGTGCTGTAA AAACTGTGGG ATGCTCCTCC CTTCTCTACC TCATGGGGGT 660

ATTGTATAAG TCCTTGCAAG AATCAGTGCA AAGATTTGCT TTAATTGTTA AGATATGATG 720

TCCCTATGGA AGCATATTGT TATTATATAA TTACATATTT GCATATGTAT GACTCCCAAA 780

TTTTCACATA AAATAGATTT TTGTAAAAAA 810

(2) INFORMATION FOR SEQ ID NO:36:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 91 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Met Lys Val Ser Ala Ala Arg Leu Ala Val Ile Leu Ile Ala Thr Ala
1 5 10 15

Leu Cys Ala Pro Ala Ser Ala Ser Pro Tyr Ser Ser Asp Thr Thr Pro
20 25 30

Cys Cys Phe Ala Tyr Ile Ala Arg Pro Leu Pro Arg Ala His Ile Lys
35 40 45

Glu Tyr Phe Tyr Thr Ser Gly Lys Cys Ser Asn Pro Ala Val Val Phe
50 55 60

Val Thr Arg Lys Asn Arg Gln Val Cys Ala Asn Pro Glu Lys Lys Trp
65 70 75 80

Val Arg Glu Tyr Ile Asn Ser Leu Glu Met Ser
85 90

(2) INFORMATION FOR SEQ ID NO:37:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1160 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

CCTCCGACAG CCTCTCCACA GGTACCATGA AGGTCTCCGC GGCACGCCTC GCTGTCATCC 60

TCATTGCTAC TGCCCTCTGC GCTCCTGCAT CTGCCTCCCC ATATTCCTCG GACACCACAC 120

CCTGCTGCTT TGCCTACATT GCCCGCCCAC TGCCCCGTGC CCACATCAAG GAGTATTTCT 180

ACACCAGTGG CAAGTGCTCC AACCCAGCAG TCGTCTTTGT CACCCGAAAG AACCGCCAAG 240

TGTGTGCCAA CCCAGAGAAG AAATGGGTTC GGGAGTACAT CAACTCTTTG GAGATGAGCT 300

AGGATGGAGA GTCCTTGAAC CTGAACTTAC ACAAATTTGC CTGTTTCTGC TTGCTCTTGT 360

CCTAGCTTGG GAGGCTTCCC CTCACTATCC TACCCCACCC GCTCCTTGAA GGGCCCAGAT 420

TCTGACCACG ACGAGCAGCA GTTACAAAAA CCTTCCCCAG GCTGGACGTG GTGGCTCAGC 480

CTTGTAATCC CAGCACTTTG GGAGGCCAAG GTGGGTGGAT CACTTGAGGT CAGGAGTTCG 540

AGACAGCCTG GCCAACATGA TGAAACCCCA TGTGTACTAA AAATACAAAA AATTAGCCGG 600

GCGTGGTAGC GGGCGCCTGT AGTCCCAGCT ACTCGGGAGG CTGAGGCAGG AGAATGGCGT 660

GAACCCGGGA GCGGAGCTTG CAGTGAGCCG AGATCGCGCC ACTGCACTCC AGCCTGGGCG 720

ACAGAGCGAG ACTCCGTCTC AAAAAAAAAA AAAAAAAAAA AAAAAATACA AAAATTAGCC 780

GCGTGGTGGC CCACGCCTGT AATCCCAGCT ACTCGGGAGG CTAAGGCAGG AAAATTGTTT 840

GAACCCAGGA GGTGGAGGCT GCAGTGAGCT GAGATTGTGC CACTTCACTC CAGCCTGGGT 900

GACAAAGTGA GACTCCGTCA CAACAACAAC AACAAAAAGC TTCCCCAACT AAAGCCTAGA 960

AGAGCTTCTG AGGCGCTGCT TTGTCAAAAG GAAGTCTCTA GGTTCTGAGC TCTGGCTTTG 1020

CCTTGGCTTT GCAAGGGCTC TGTGACAAGG AAGGAAGTCA GCATGCCTCT AGAGGCAAGG 1080

AAGGGAGGAA CACTGCACTC TTAAGCTTCC GCCGTCTCAA CCCCTCACAG GAGCTTACTG 1140

GCAAACATGA AAAATCGGGG 1160

(2) INFORMATION FOR SEQ ID NO:38:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 97 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

Met Arg Ile Ser Ala Thr Leu Leu Cys Leu Leu Leu Ile Ala Ala Ala
1 5 10 15

Phe Ser Ile Gln Val Trp Ala Gln Pro Asp Gly Pro Asn Ala Ser Thr
20 25 30

Cys Cys Tyr Val Lys Lys Gln Lys Ile Pro Lys Arg Asn Leu Lys Ser
35 40 45

Tyr Arg Arg Ile Thr Ser Ser Arg Cys Pro Trp Glu Ala Val Ile Phe
50 55 60

Lys Thr Lys Lys Gly Met Glu Val Cys Arg Glu Ala His Gln Lys Trp
65 70 75 80

Val Glu Glu Ala Ile Ala Tyr Leu Asp Met Lys Thr Pro Thr Pro Lys
85 90 95

Pro

(2) INFORMATION FOR SEQ ID NO:39:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 593 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

ACTGAAGCCA GCTCTCTCAC TCTCTTTCTC CACCATGAGG ATCTCTGCCA CGCTTCTGTG 60

CCTGCTGCTC ATAGCCGCTG CTTTCAGCAT CCAAGTGTGG GCCCAACCAG ATGGGCCCAA 120

TGCATCCACA TGCTGCTATG TCAAGAAACA AAGATCCCC AAGAGGAATC TCAAGAGCTA 180

CAGAAGGATC ACCAGTAGTC GGTGTCCCTG GGAAGCTGTT ATCTTCAAGA CAAAGAAGGG 240

CATGGAAGTC TGTCGTGAAG CCCATCAGAA GTGGGTCGAG GAGGCTATAG CATACTTAGA 300

CATGAAAACC CCAACTCCAA AGCCTTGAAG AAATGTGCCT GAACAGAAAC CAACCTAGGA 360

GCCAAGAAGC AAAAATTCCT CACCGCTGTT CTTTCTGAGA ACTGTTGATG AAATGTGTTG 420

ATCACGGTCC TAAGGGATAG GAGCTGTCTG TAGGAATGTG AAACAGTCAC GCCTAAGGAA 480

TGGTCTTTAA GTTATTAATA TTTTTATTTA ATTAGCCATG TACTTTGGTG TGATTTGAAT 540

GTAAAGCTCT GGAGACCTCA TGTCACTTTA ACATTGTGTT AGCTGCAGAA TTC 593

(2) INFORMATION FOR SEQ ID NO:40:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 72 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

Asp Ser Val Ser Ile Phe Ile Thr Cys Cys Phe Asn Val Ile Asn Arg
1 5 10 15

Lys Ile Pro Ile Gln Arg Leu Glu Ser Tyr Thr Arg Ile Thr Asn Ile
20 25 30

Gln Cys Pro Lys Glu Ala Val Ile Phe Lys Thr Gly Lys Glu Val Cys
35 40 45

Ala Asp Pro Lys Glu Arg Trp Val Arg Asp Ser Met Lys His Lys Asp
50 55 60

Gln Ile Phe Gln Asn Leu Lys Pro
65 70

(2) INFORMATION FOR SEQ ID NO:41:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 15 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

Asn Leu Ala Lys Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Cys
1 5 10 15

(2) INFORMATION FOR SEQ ID NO:42:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 32 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

Cys Asn Gln Val Glu Val Ile Ala Thr Leu Lys Asp Gly Arg Lys Ile
1 5 10 15

Cys Leu Asp Pro Asp Ala Pro Arg Ile Lys Lys Ile Val Gln Lys Lys
20 25 30

(2) INFORMATION FOR SEQ ID NO:43:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 96 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

TGCAACCAAG TCGAAGTGAT AGCCACACTG AAGGATGGGA GGAAAATCTG CCTGGACCCA 60

GATGCTCCCA GAATCAAGAA AATTGTACAG AAAAAA 96

CLAIMS

We claim:

1. A method of screening for AHA compounds comprising the steps of:
   a) contacting a compound with radiolabeled heparin/heparan sulfate and heparanase;
   b) maintaining the compounds in contact with the radiolabeled heparin/heparan sulfate and heparanase for a time and under such conditions sufficient to allow inhibition of heparanase activity;
   c) detecting inhibition of heparanase activity (a compound that gives 50% inhibition at a concentration of 1 μM or less); and
   d) selecting compounds that inhibit heparanase activity.

2. A method according to claim 1 wherein the heparanase is recombinant.

3. A heparanase having an isoelectric point of less than 5.5 and possessing activity greater than 20 units heparanase activity per μg protein.

4. A heparanase according to Claim 3, having an isoelectric point of about 4.8 - 5.1.

5. A heparanase purified to apparent homogeneity, as in claim 3, prepared in the presence of reducing conditions and activated with transglutaminase, having an amino acid sequence selected from the group consisting of SEQ. ID NO: 1, SEQ. ID NO: 3, SEQ. ID NO: 5 or SEQ. ID NO: 7.

6. A heparanase purified to apparent homogeneity, as in claim 3, prepared in the presence of reducing conditions, having an amino acid sequence of SEQ. ID NO: 1.

7. A heparanase purified to apparent homogeneity, as in claim 3, prepared in the presence of reducing conditions and activated with transglutaminase, having an amino acid sequence of SEQ. ID NO: 3.

8. A heparanase according to Claim 4, purified to apparent homogeneity, prepared in the presence of reducing conditions and activated with transglutaminase.

9. A heparanase, as in claim 3, prepared by recombinant means, activated with transglutaminase and having an amino acid sequence selected from the group consisting of SEQ.

ID NO: 1, SEQ. ID NO: 3, SEQ. ID NO: 5 or SEQ. ID NO: 7.

10. A heparanase, as in claim 3, prepared by recombinant means, activated with transglutaminase and having an amino acid sequence of SEQ. ID NO: 1.

11. A heparanase, as in claim 3, prepared by recombinant means, activated with transglutaminase and having an amino acid sequence of SEQ. ID NO: 3.

12. A heparanase purified to apparent homogeneity, as in claim 3, prepared in the presence of reducing conditions and activated with transglutaminase, and having an amino acid sequence selected from the group consisting of SEQ. ID. NO: 12, SEQ. ID. NO: 14; SEQ. ID. NO: 16, SEQ. ID. NO: 18, SEQ. ID. NO: 20, SEQ. ID. NO: 22 and SEQ. ID. NO: 24.

13. A heparanase purified to apparent homogeneity, as in claim 3, prepared in the presence of reducing conditions and activated with transglutaminase, and having an amino acid sequence selected from the group consisting of SEQ. ID. NO: 26, SEQ. ID. NO: 28, SEQ. ID. NO: 30, SEQ. ID. NO: 32; SEQ. ID. NO: 34, SEQ. ID. NO: 36, SEQ. ID. NO: 38 and SEQ.ID. NO: 40.

14. A method according to claim 1 wherein the heparanase is purified to apparent homogeneity, prepared in the presence of reducing conditions and activated with transglutaminase, and having an amino acid sequence selected from the group consisting of SEQ. ID. NO: 12, SEQ. ID. NO: 14; SEQ. ID. NO: 16, SEQ. ID. NO: 18, SEQ. ID. NO: 20, SEQ. ID. NO: 22 and SEQ. ID. NO: 24.

15. A method according to claim 1 wherein the heparanase is purified to apparent homogeneity, prepared in the presence of reducing conditions and activated with transglutaminase, and having an amino acid sequence selected from the group consisting of SEQ. ID. NO: 26, SEQ. ID. NO: 28, SEQ. ID. NO: 30, SEQ. ID. NO: 32; SEQ. ID. NO: 34, SEQ. ID. NO: 36, SEQ. ID. NO: 38 and SEQ. ID. NO: 40.

16. A method according to claim 1 wherein the heparanase is purified to apparent homogeneity, prepared in the presence of reducing conditions and activated with transglutaminase, and having an amino acid sequence selected from the group consisting of SEQ. ID NO: 1, SEQ. ID NO: 3, SEQ. ID NO: 5 or SEQ. ID NO: 7.

17. A peptide having an amino acid sequence of SEQ. ID. NO: 42.

INTERNATIONAL SEARCH REPORT

Inter nal Application No
PCT/US 94/08207

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12Q1/34 C12N9/24 C12N9/96 C07K14/47

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 C12Q C12N A61K C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	ANALYTICAL BIOCHEMISTRY, vol.157, no.1, 15 August 1986, NEW YORK US pages 162 - 171 MOTOWO NAKAJIMA ET AL. 'A Solid-Phase Substrate of Heparanase: Its Application to Assay of Human Melanoma for Heparan Sulfate Degradative Activity' see the whole document ---	1,2
A	JOURNAL OF BIOLOGICAL CHEMISTRY, vol.259, no.4, 25 February 1984, BALTIMORE, MD US pages 2283 - 2290 MOTOWO NAKAJIMA ET AL. 'Metastatic Melanoma Cell Heparanase' see the whole document	1,2
A	see page 2289, left column, line 63 - right column, line 31 --- -/--	3,4

[X] Further documents are listed in the continuation of box C.

[X] Patent family members are listed in annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
9 December 1994	21. 12. 94
Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax (+31-70) 340-3016	Authorized officer Döpfer, K-P

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INTERNATIONAL SEARCH REPORT

Int onal Application No
PCT/US 94/08207

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO,A,91 02977 (HADASSAH MEDICAL ORGANIZATION) 7 March 1991 cited in the application see page 23, paragraph 6.1.5 - page 26, paragraph 6.1.7; claims 1-3	3-16
A	FASEB JOURNAL, vol.5, no.15, December 1991, BETHESDA, MD US pages 3071 - 3076 CHARLES S. GREENBERG ET AL. 'Transglutaminases: multifunctional cross-linking enzymes that stabilize tissues' cited in the application see the whole document	3-16
A	JOURNAL OF BIOLOGICAL CHEMISTRY, vol.265, no.28, 5 October 1990, BALTIMORE, MD US pages 17180 - 17188 ELEONORA CORDELLA-MIELE ET AL. 'A Novel Transglutaminase-mediated Post-translational Modification of Phospholipase A2 Dramatically Increases Its Catalytic Activity' cited in the application see the whole document	3-16
A	WO,A,93 09794 (UNIVERSITY OF PENNSYLVANIA) 27 May 1993 see especially Sequence Desription: SEQ ID NO: 1 see page 17, line 1 - line 16	17

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INTERNATIONAL SEARCH REPORT

Information on patent family members

Int onal Application No
PCT/US 94/08207

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
WO-A-9102977	07-03-91	AU-B-	654804	24-11-94
		AU-A-	6336490	03-04-91
		CA-A-	2065744	24-02-91
		EP-A-	0487627	03-06-92
		JP-T-	5504047	01-07-93
		US-A-	5362641	08-11-94
WO-A-9309794	27-05-93	AU-A-	3071492	15-06-93